

INSECTICIDE RESISTANCE IN NORTH AMERICAN WEST NILE VECTORS.

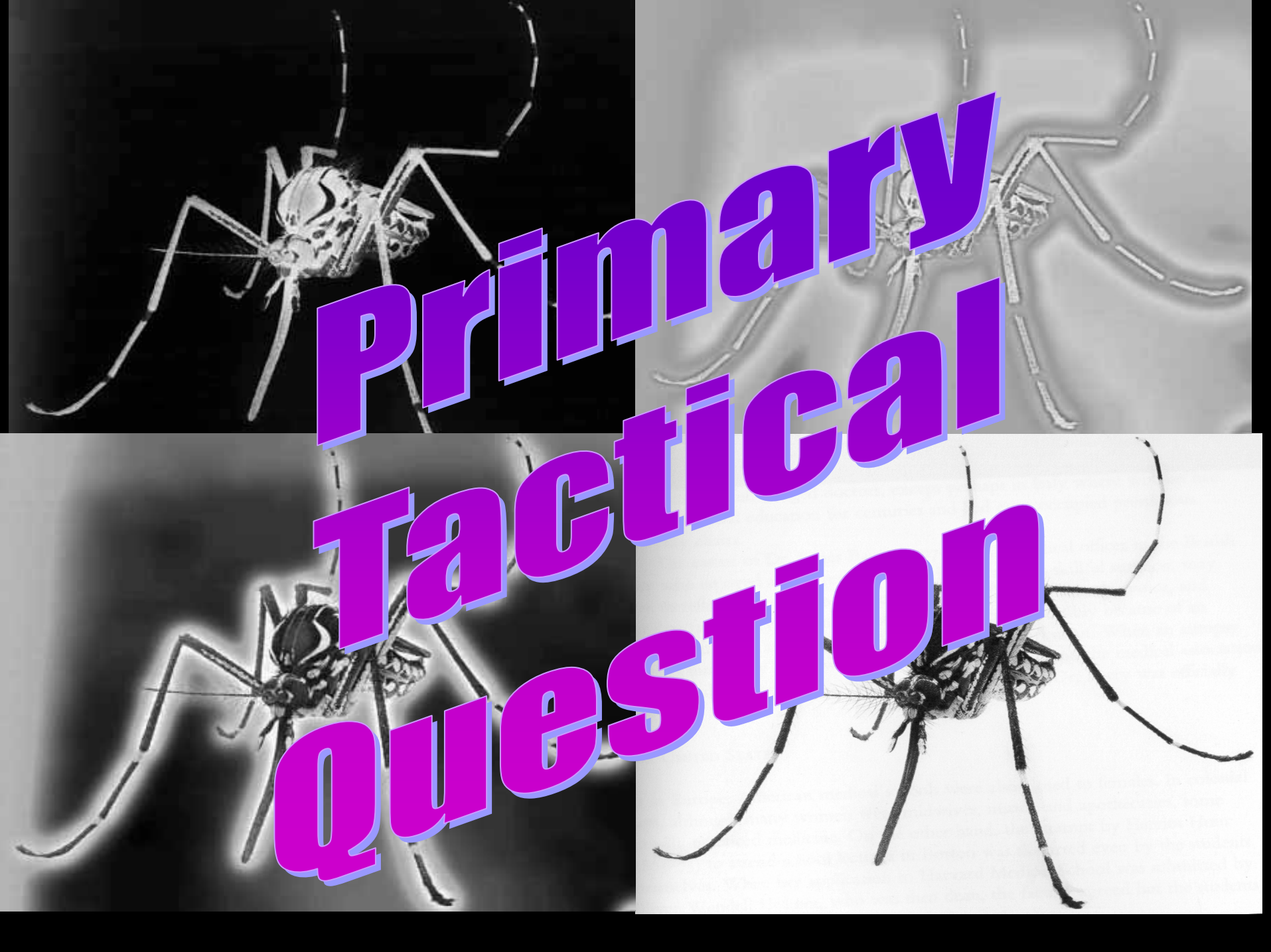
WILLIAM G. BROGDON, PH.D.

ATHENS, GEORGIA OCTOBER 18TH, 2007

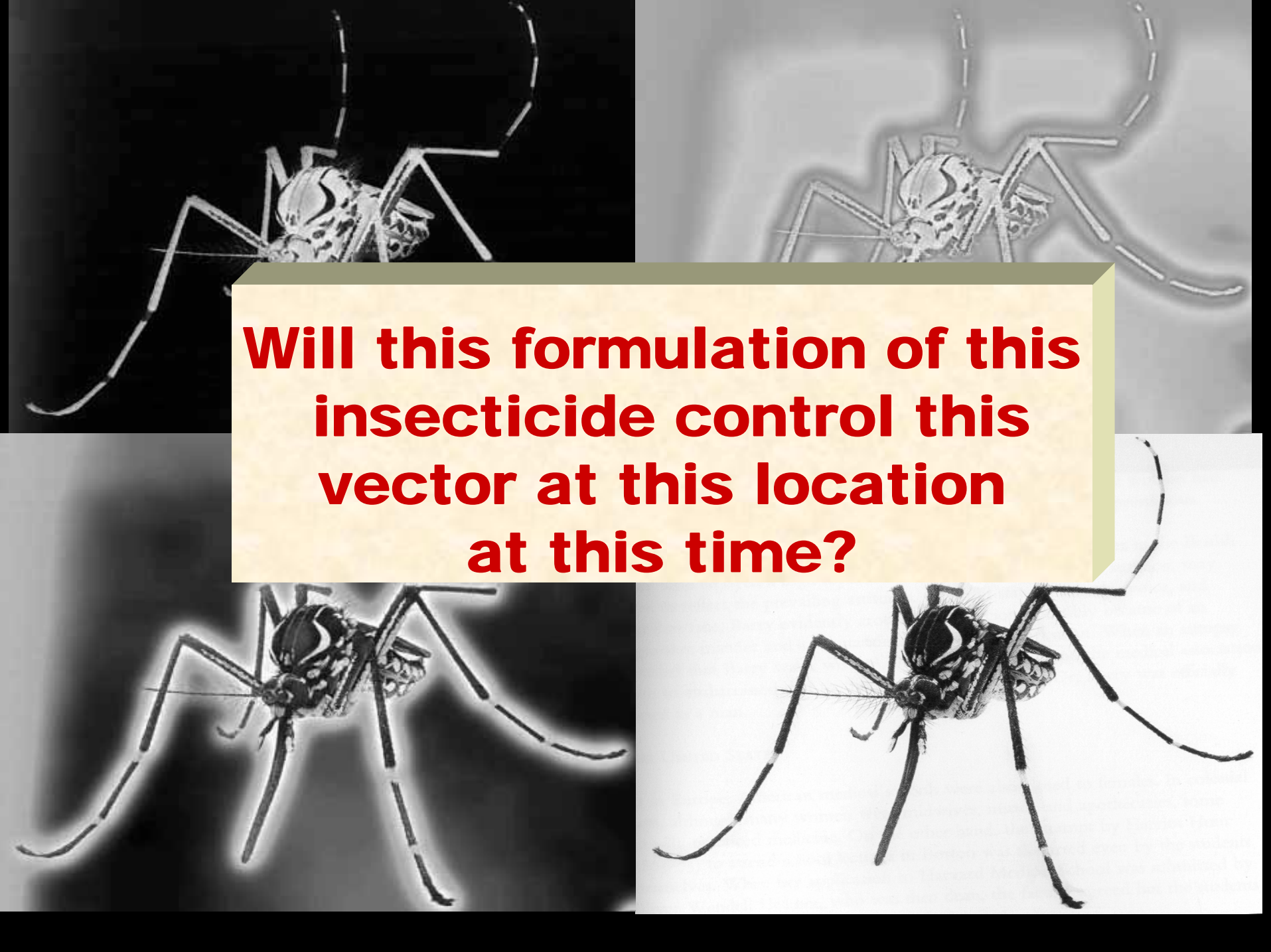




THE MOST PRACTICAL METHOD FOR PROVING THE PRESENCE OF RESISTANCE IS A GOOD BIOASSAY.



**Primary
Tactical
Question**



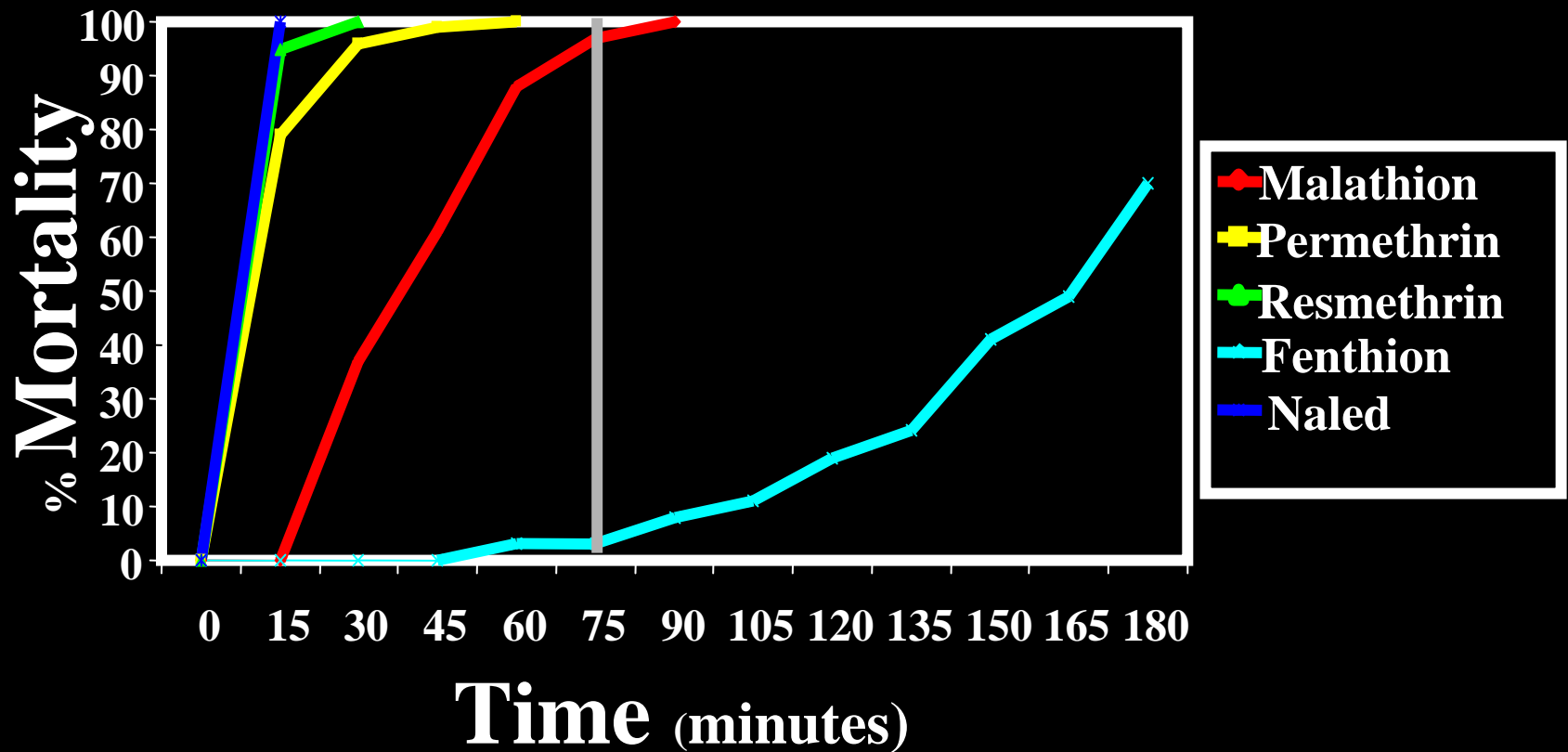
Will this formulation of this insecticide control this vector at this location at this time?



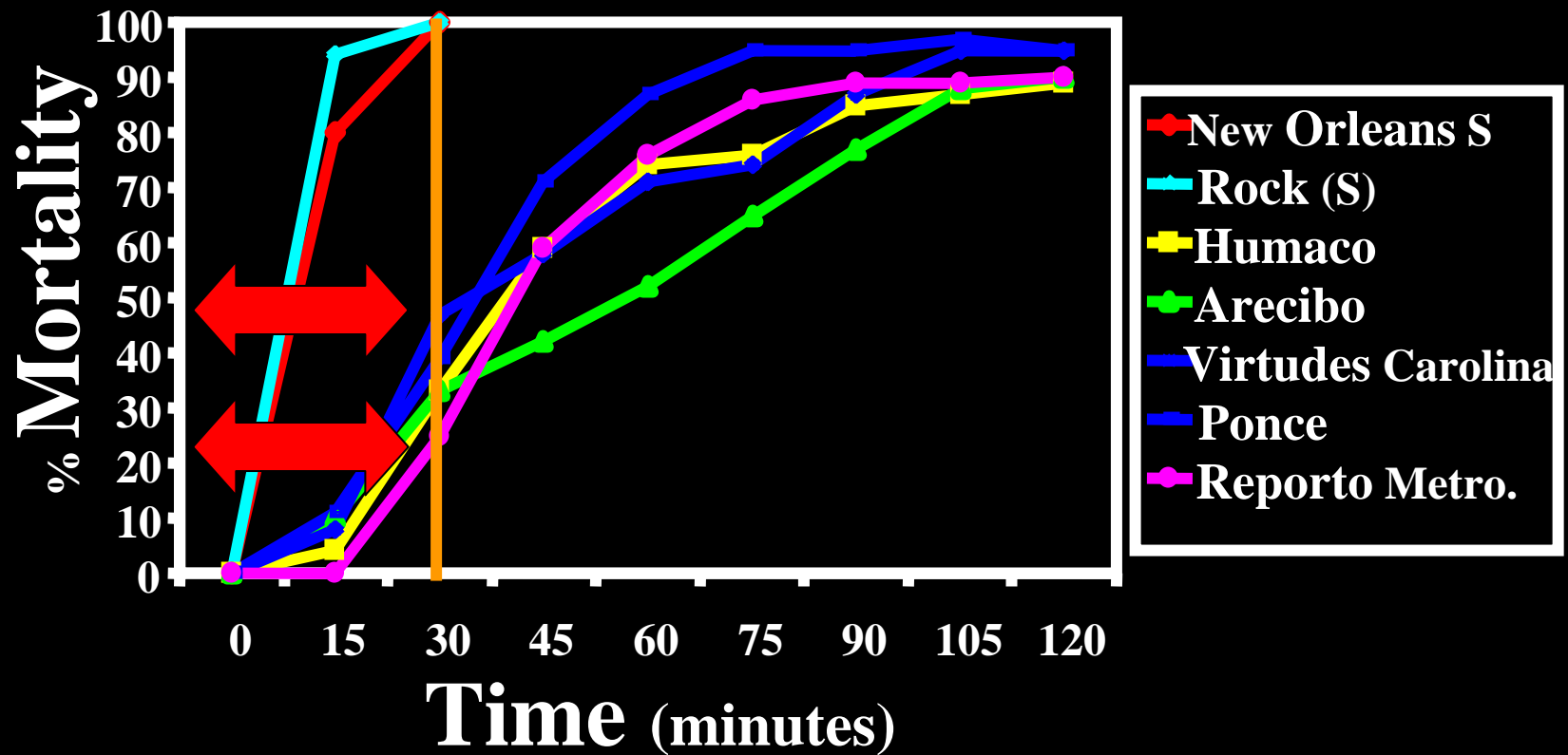
If not,
what do I do now?



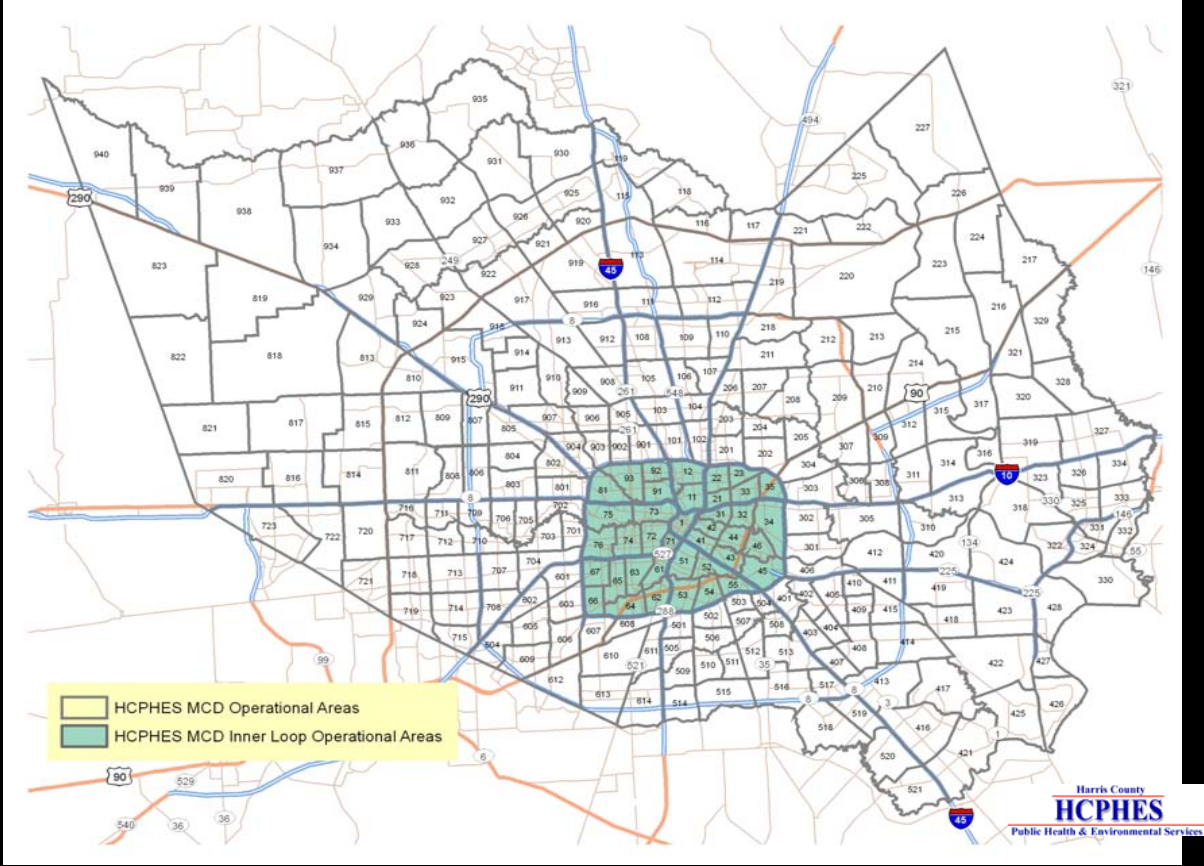
Bottle bioassays run on *Culex nigripalpus* from Indian River, Florida



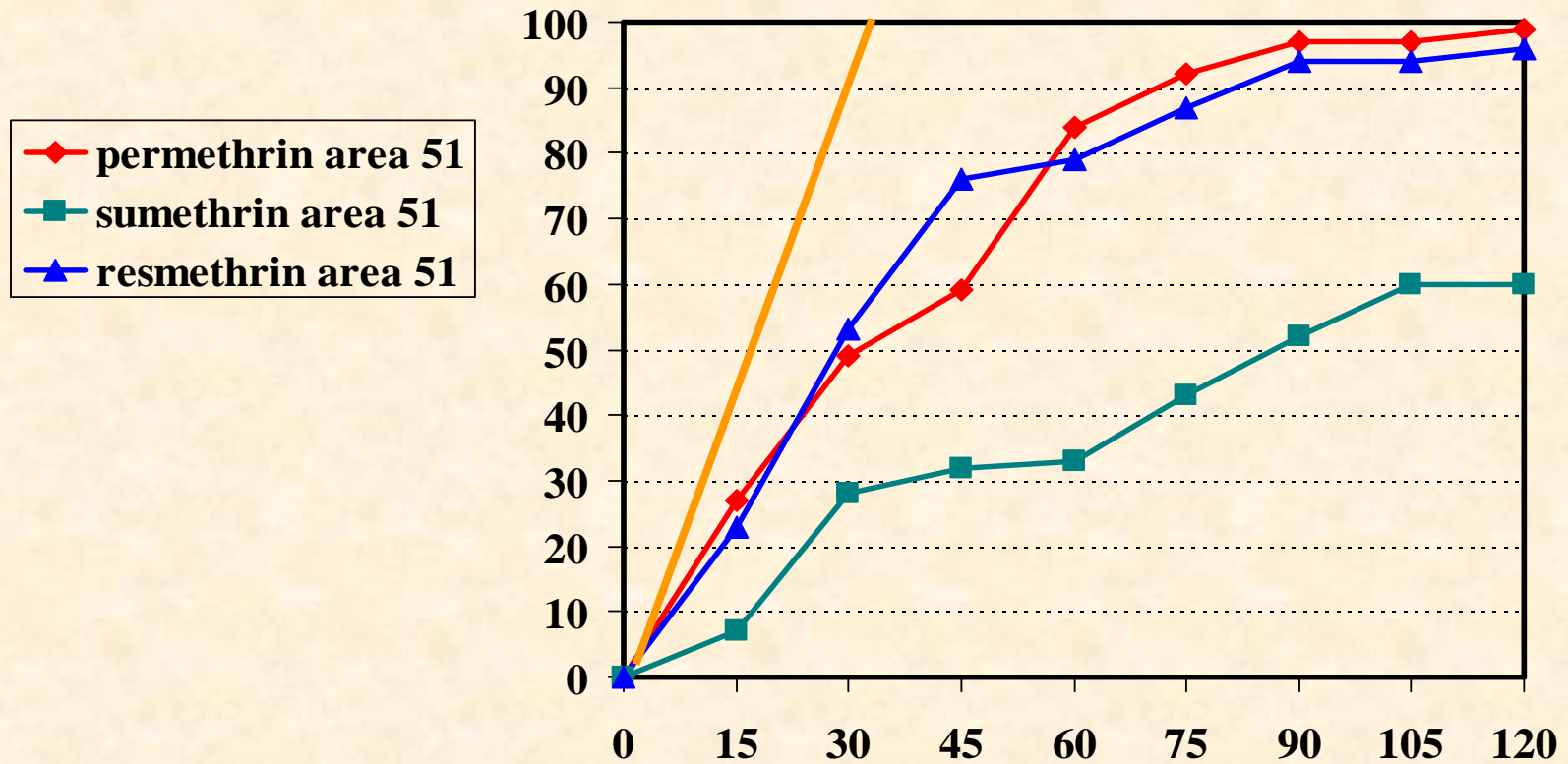
1999 Puerto Rico *Aedes aegypti* assayed with Permethrin

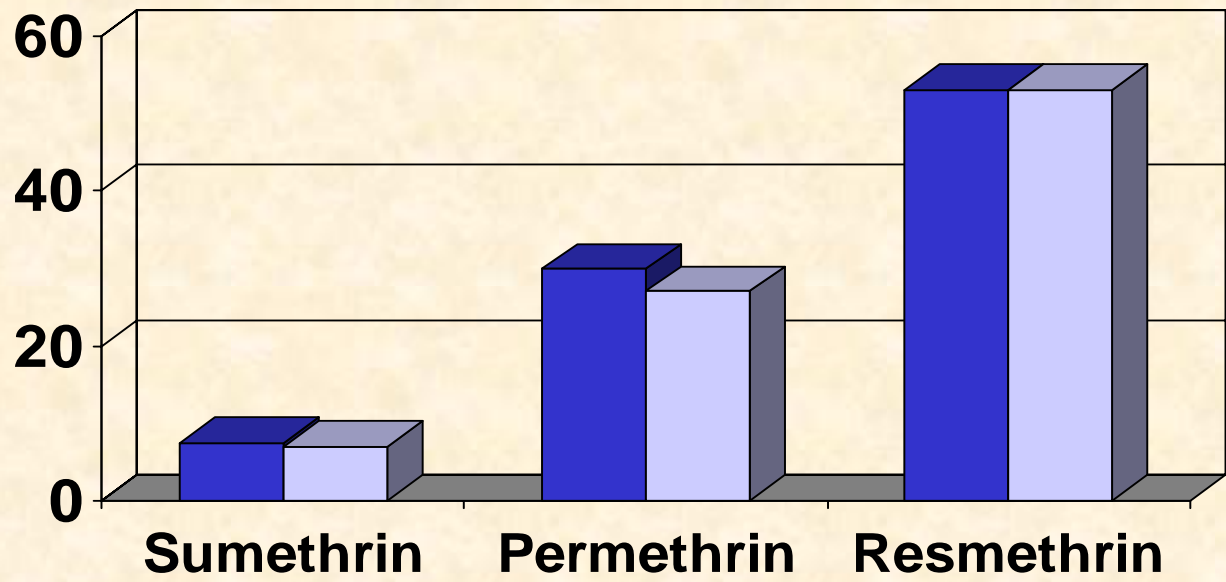


Mosquito Control Operational Areas



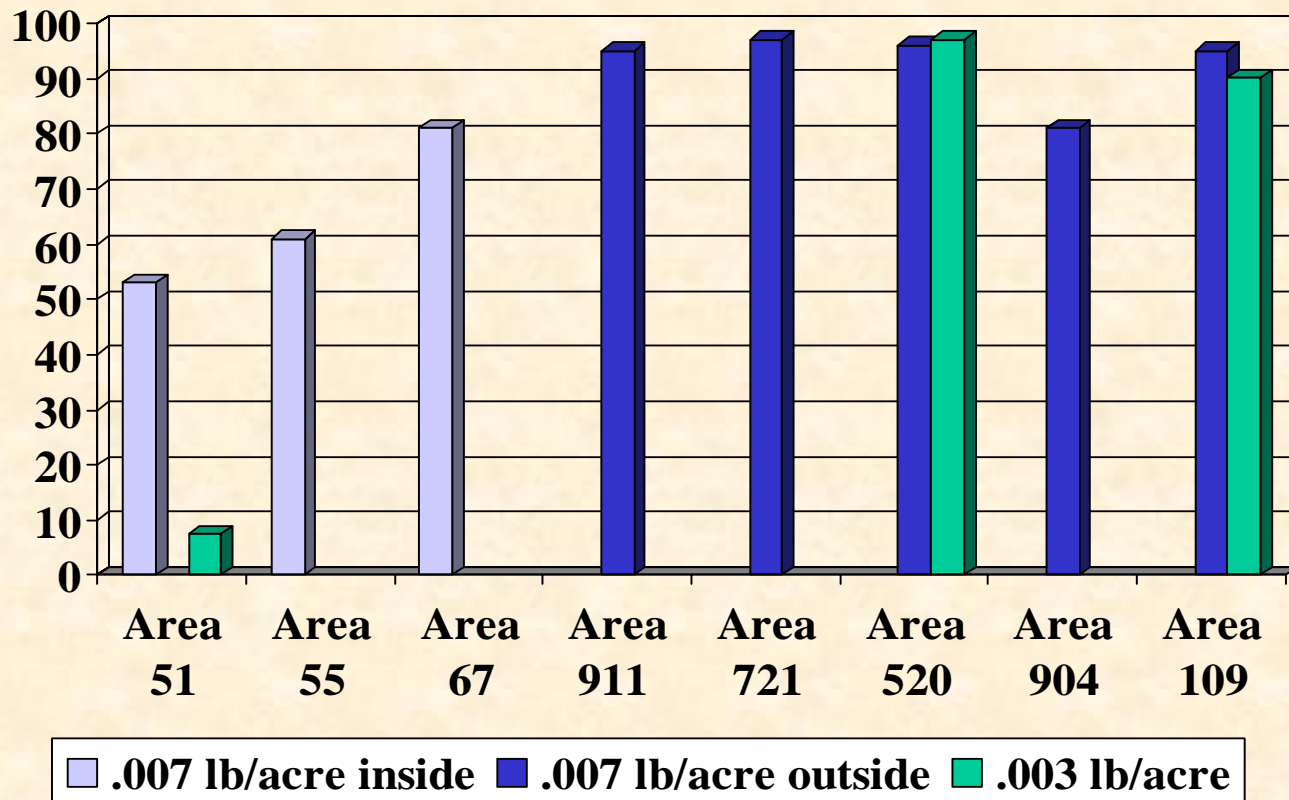
Area 51 bottle assay results

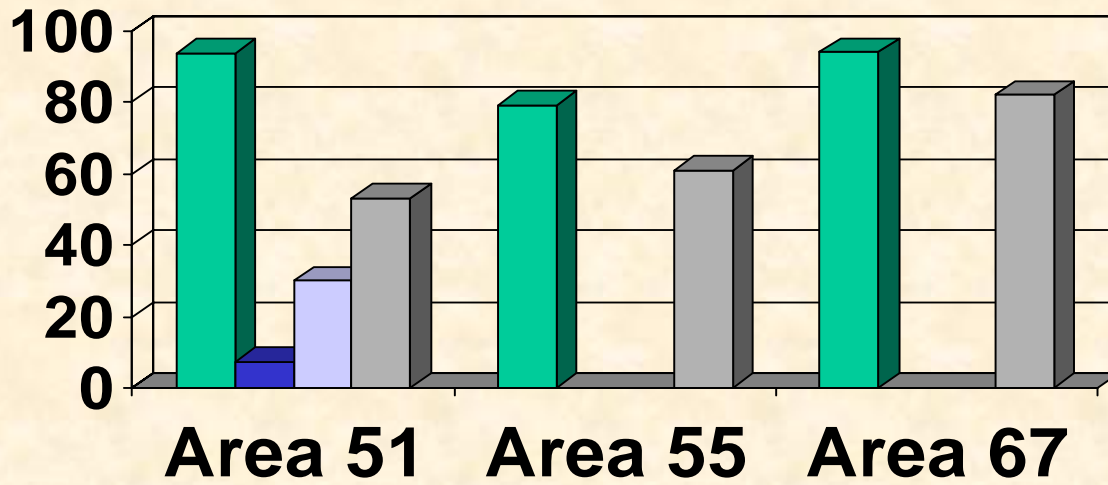




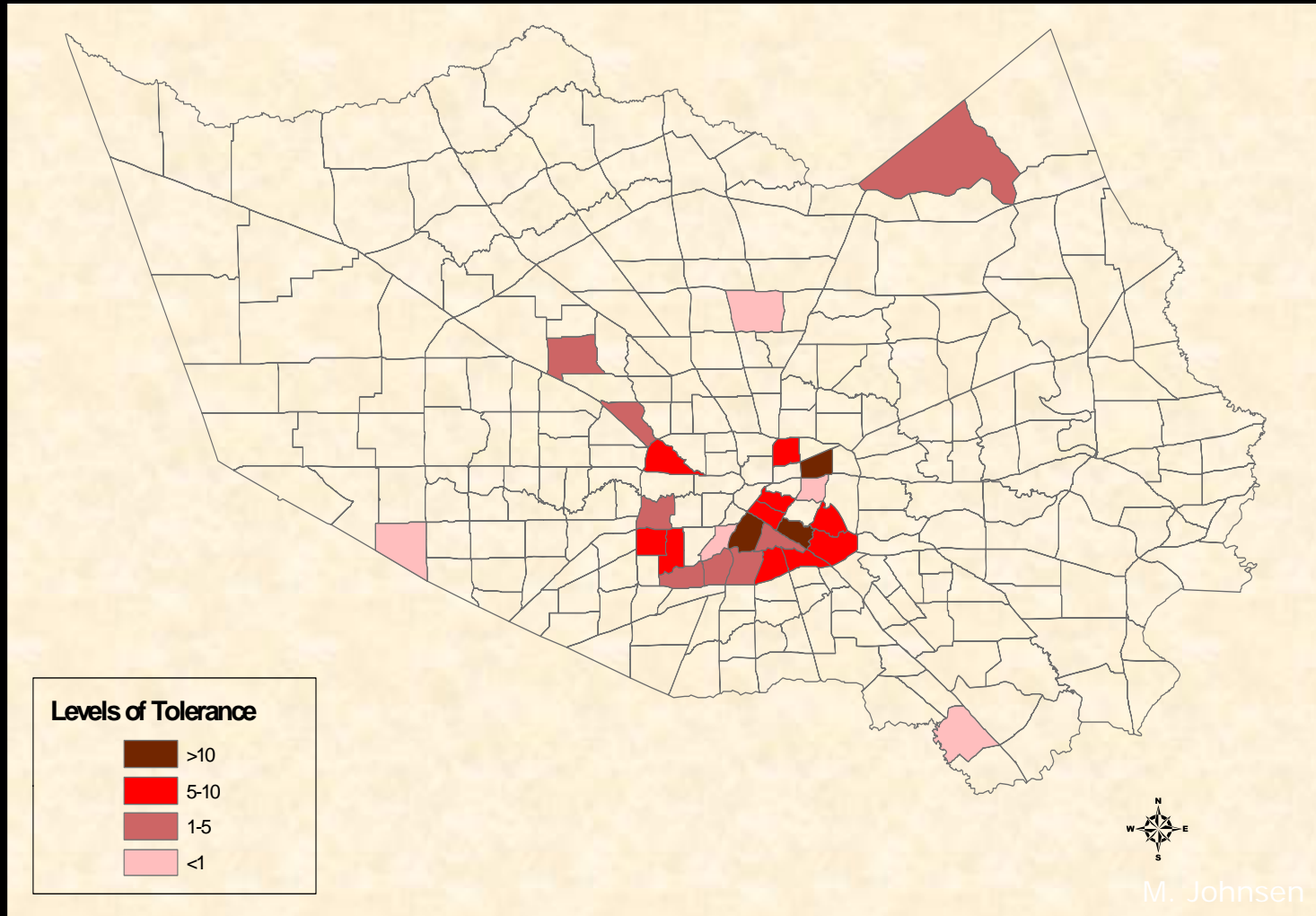
Field Tests **Bottle Tests**

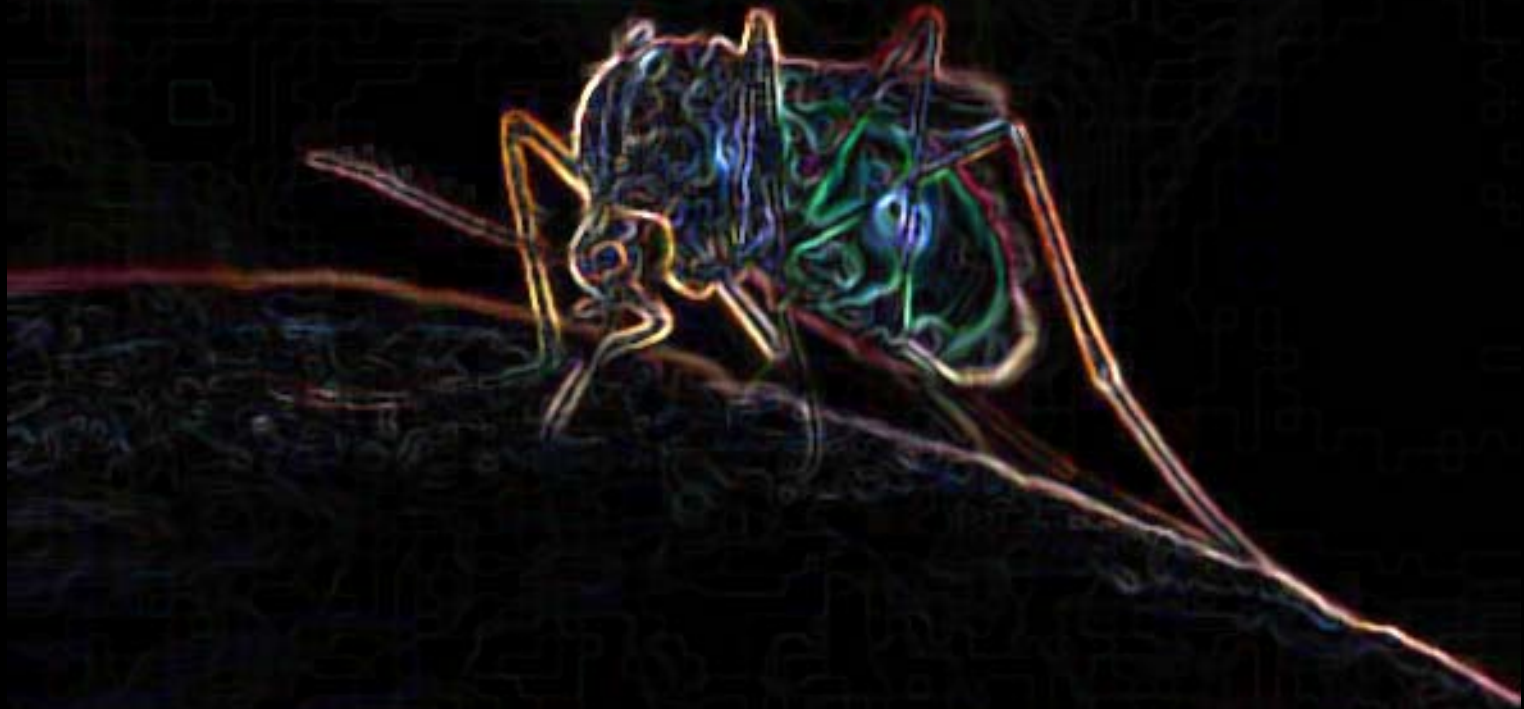
Percent mortality in feral *Culex quinquefasciatus* exposed to .003 or .007 lb/acre Resmethrin





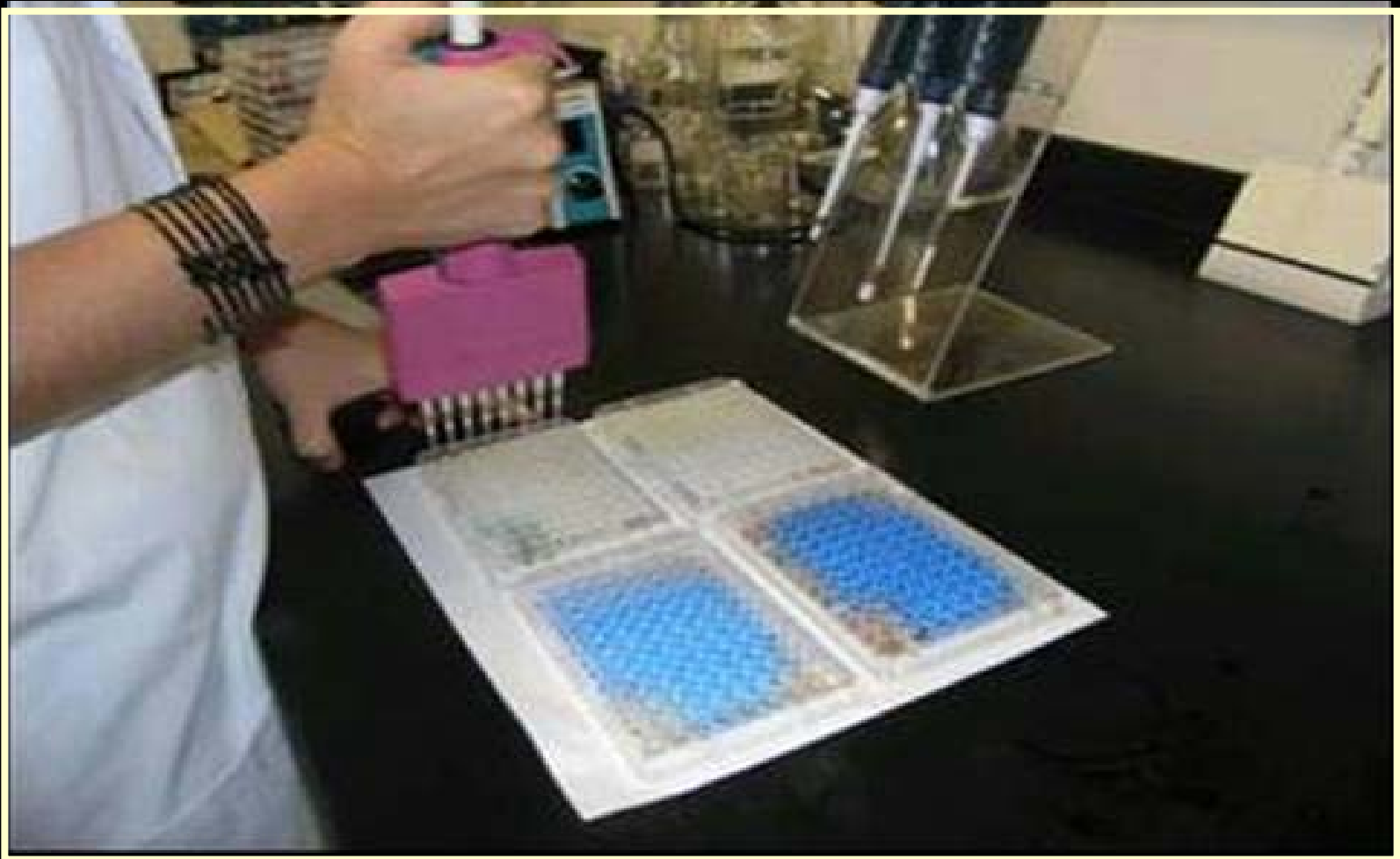
Levels of Resistance to resmethrin in *Culex quinquefasciatus* from Selected Operational Areas in Harris County, Texas





BIOCHEMICAL ASSAYS

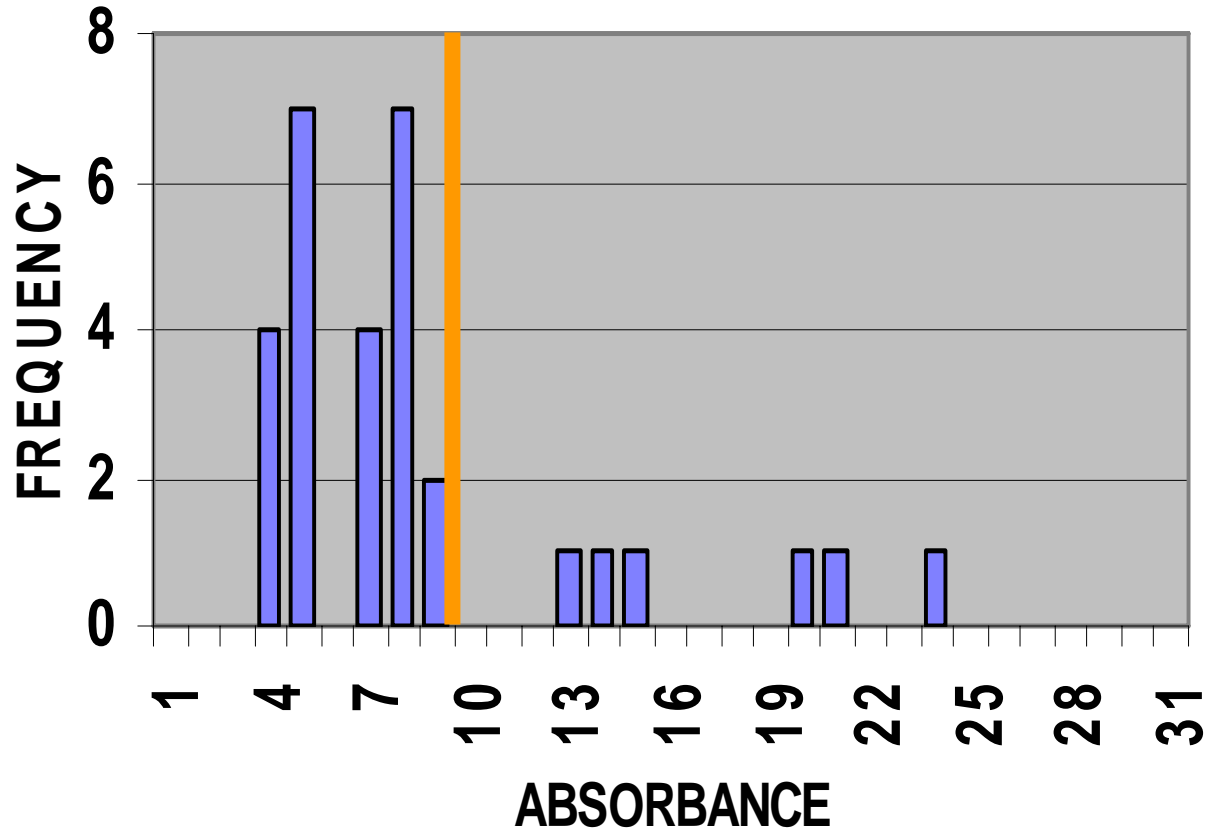
MOLECULAR ASSAYS



**MEASURE EXPRESSED PROTEIN LEVELS
MULTIPLEXED
FIVE RESISTANCE ENZYMES / INSECT**

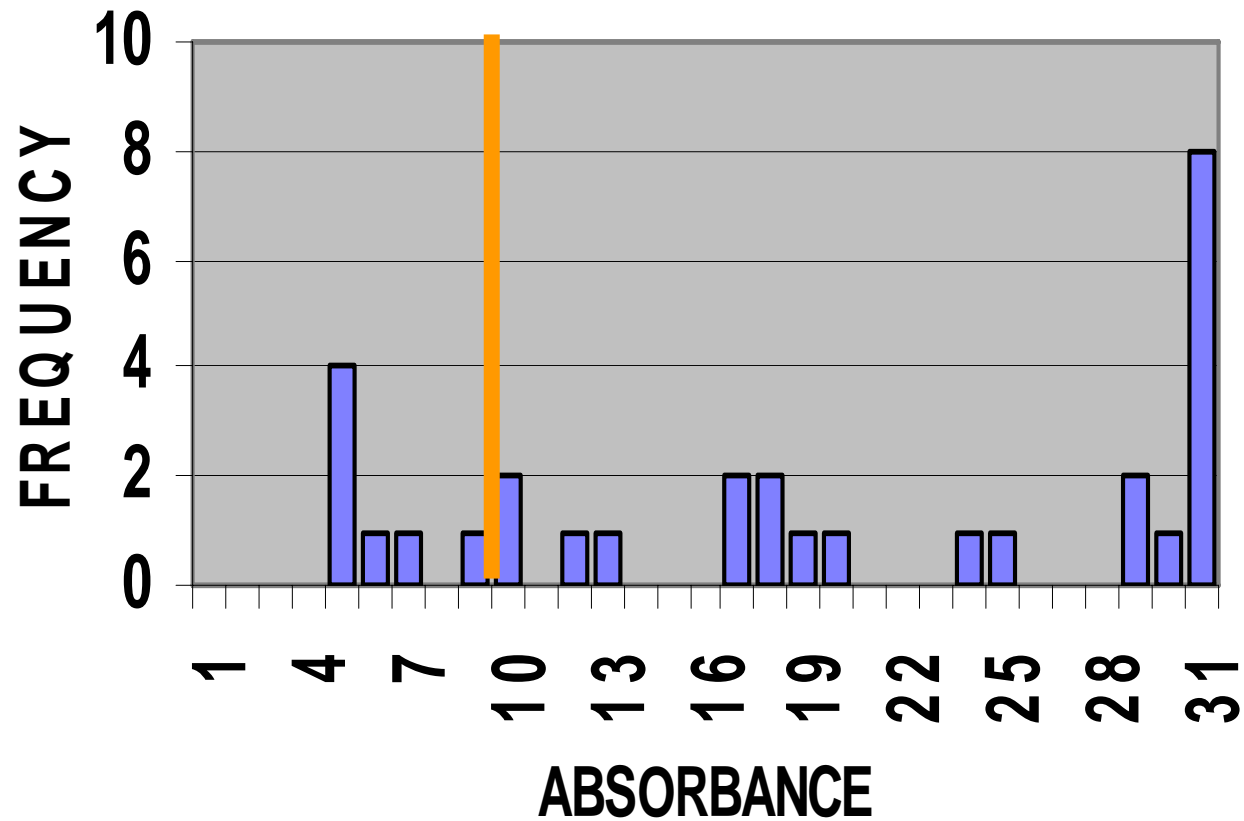
CULEX PIPIENS ESTERASE

NORTON, PORTAGE COUNTY, OHIO



CULEX PIPIENS ESTERASE

COVENTRY, SUMMIT COUNTY, OHIO





MOLECULAR ANALYSIS



**BECAUSE OF THE EXPENSE, MOLECULAR ASSAYS
MUST BE MULTIPLEXED TO ALLOW THE REQUIRED
INFORMATION TO BE OBTAINED FROM EACH INSECT.**

**THE OVERWHELMING CHALLENGE IS TO MAKE THIS
PRACTICAL.**

Direct Sequencing of Cloned Genes

Microarrays

Real Time Fluorescence PCR

MALDI-TOF Mass Spectrometry

Luminex





FLUORESCENCE PCR ALLOWS MEASUREMENT OF :



GENE EXPRESSION - QUANTITATIVE RTPCR.

OXIDASE RESISTANCE



COPY NUMBER - QUANTITATIVE PCR.

ESTERASE GENE COPY NUMBERS



POINT MUTATIONS – KDR POINT MUTATIONS



MELTING POINT ANALYSIS.



EXTREME MULTIPLEXING (x 100)

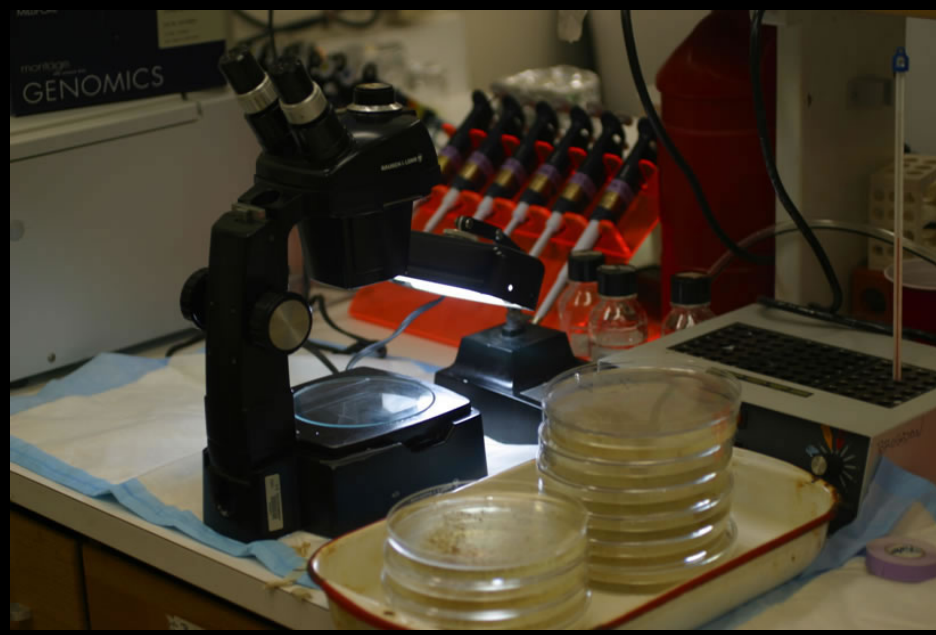
ALL RESISTANCE MECHANISMS

SPECIES ID – VECTOR

SPECIES/TYPE – PARASITE OR VIRUS

MULTIPLE SNP DATA – VECTOR

MULTIPLE SNP DATA – PATHOGEN





Mosquito collections



CURRENT INFORMATION FROM COLLECTIONS



**IDENTIFICATION OF DISEASE
POSITIVE COLLECTIONS.**

46% OF CULEX POOLS ARE “ SP. ”

POTENTIAL ADDITIONAL INFORMATION FROM COLLECTIONS



SPECIES IDENTIFICATION

RESISTANCE MECHANISM(S)

**CORRELATION OF ID, RESISTANCE
DATA, DISEASE POSITIVES**

POPULATION MARKERS (SNPs)

U.S. ARMY CENTER FOR HEALTH PROMOTION AND PREVENTATIVE MEDICINE (CHPPM)



CHPPM – North	Fort Meade Alexandra Spring
CHPPM – South	Fort McPherson Denny Kuhr
CHPPM – West	Fort Lewis Miguel Quintana

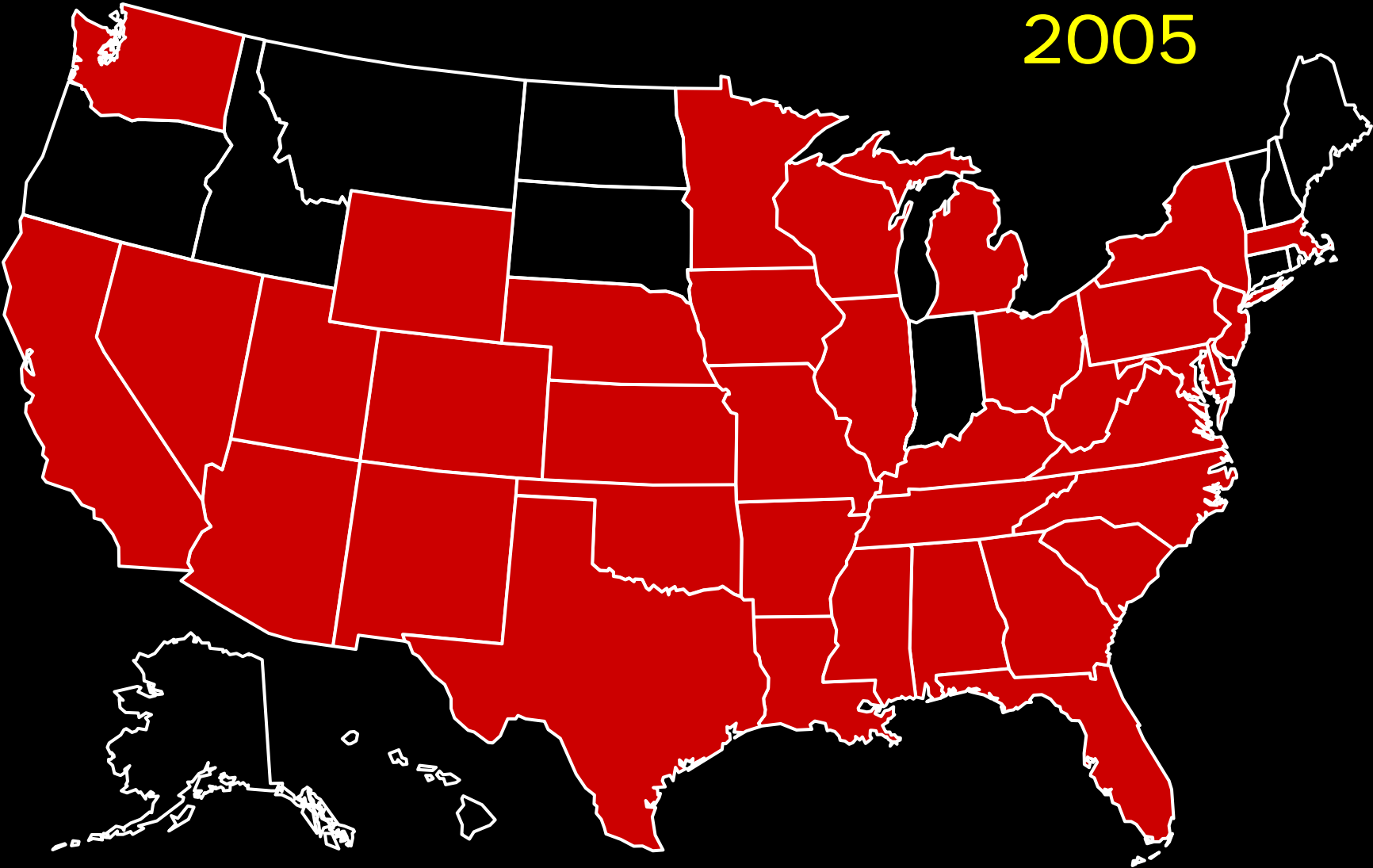


**COLLECTIONS ARE STORED
IN OUR FREEZER FACILITY**



**AT PRESENT: >55,000 COLLECTIONS
5-12 YEAR SERIES FOR MANY SITES**

2005





CULEX PIPIENS

CULEX QUINQUEFASCIATUS

CULEX MOLESTUS

CULEX RESTUANS

CULEX SALINARIUS

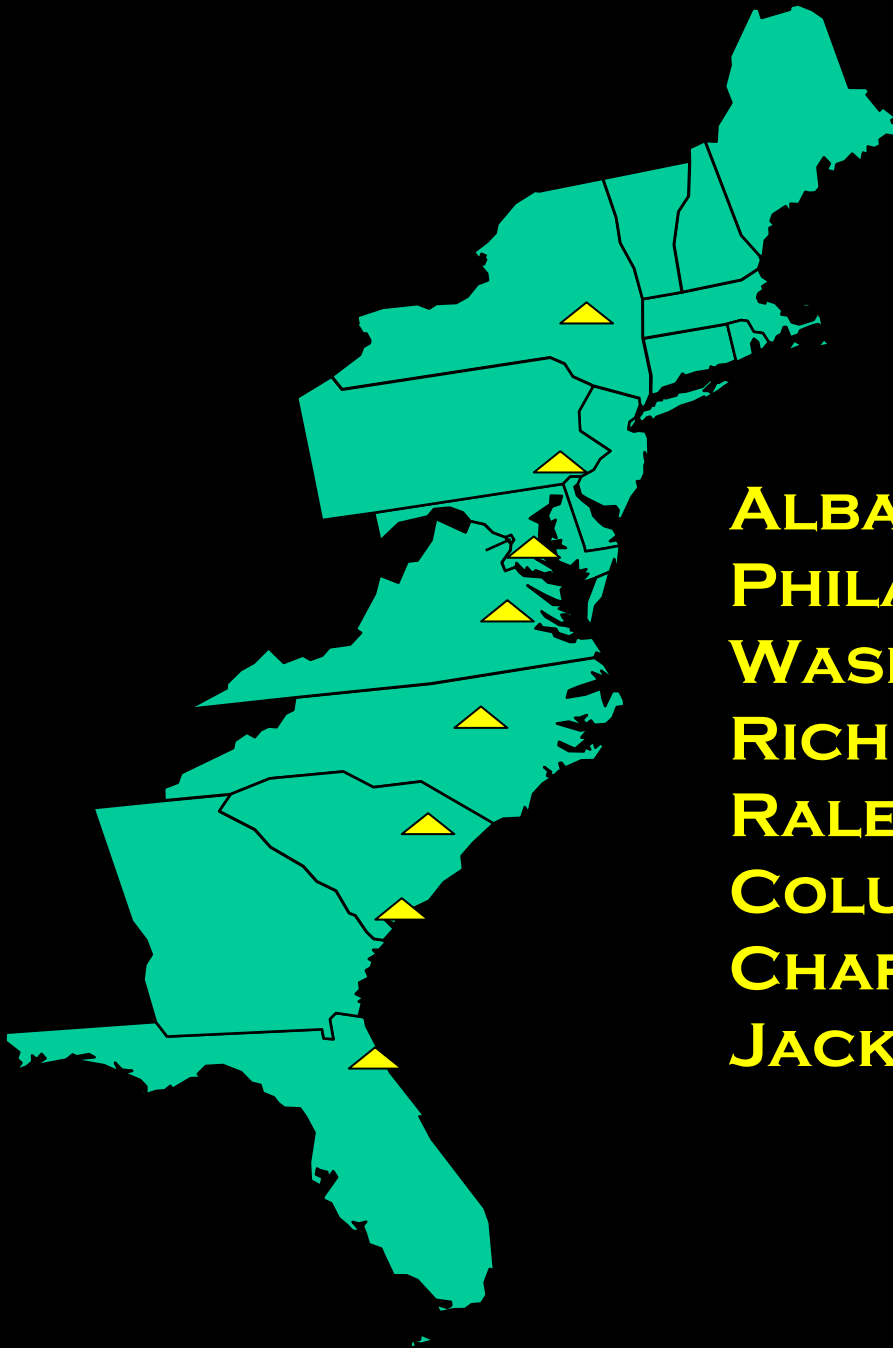
CULEX NIGRIPALPUS

CULEX TARSALIS



LAURA KRAMER, JOE VINEIS
STATE UNIVERSITY OF NEW YORK, ALBANY

DENNIS WHITE
NEW YORK DEPARTMENT OF HEALTH



ALBANY, NEW YORK

PHILADELPHIA, PENNSYLVANIA

WASHINGTON, D.C.

RICHMOND, VIRGINIA

RALEIGH, NORTH CAROLINA

COLUMBIA, SOUTH CAROLINA

CHARLESTON, SOUTH CAROLINA

JACKSONVILLE, FLORIDA

THREE TYPES OF RESISTANCE GENES IN CULEX.



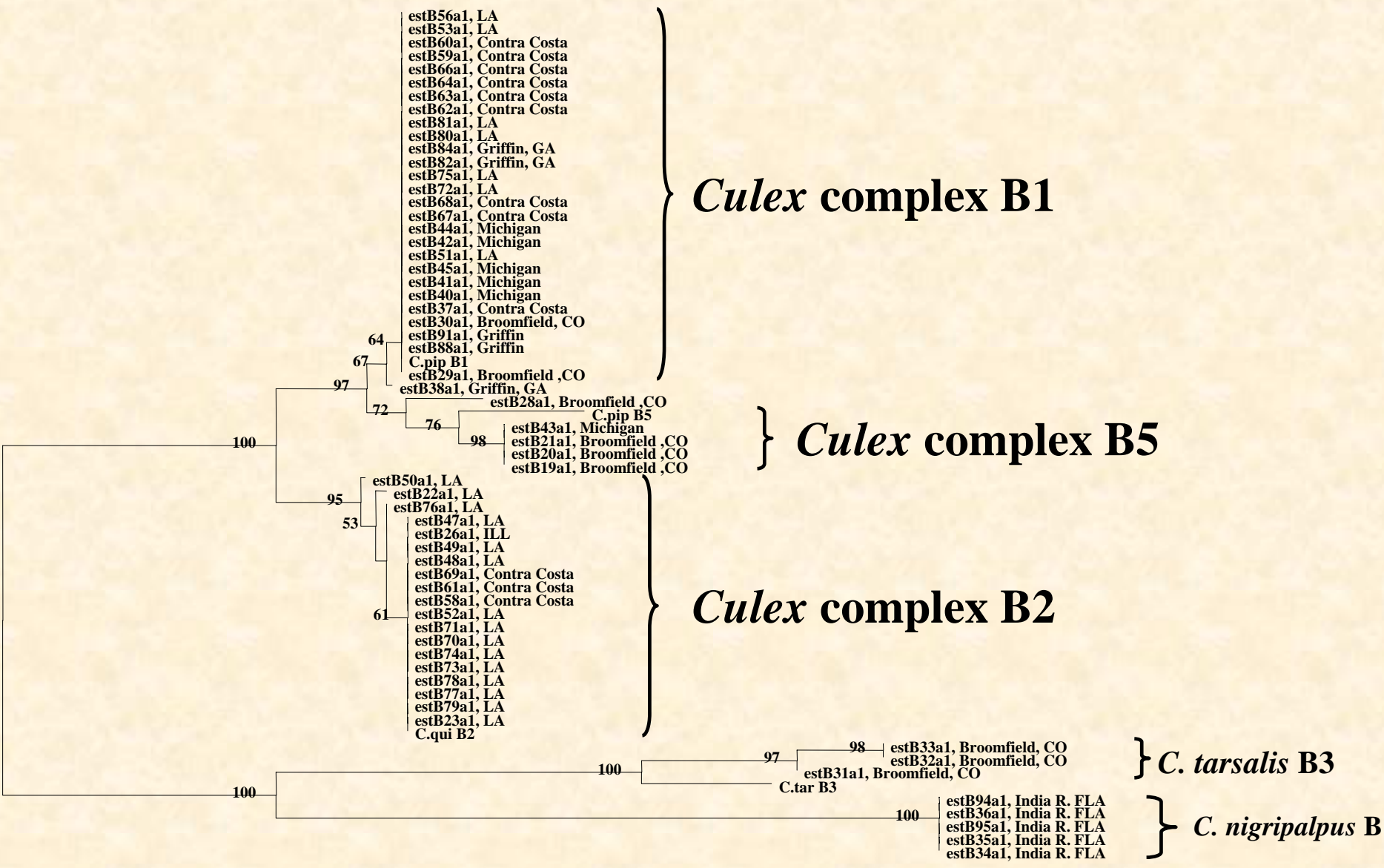
MULTIPLE COPY GENES (ESTERASES)

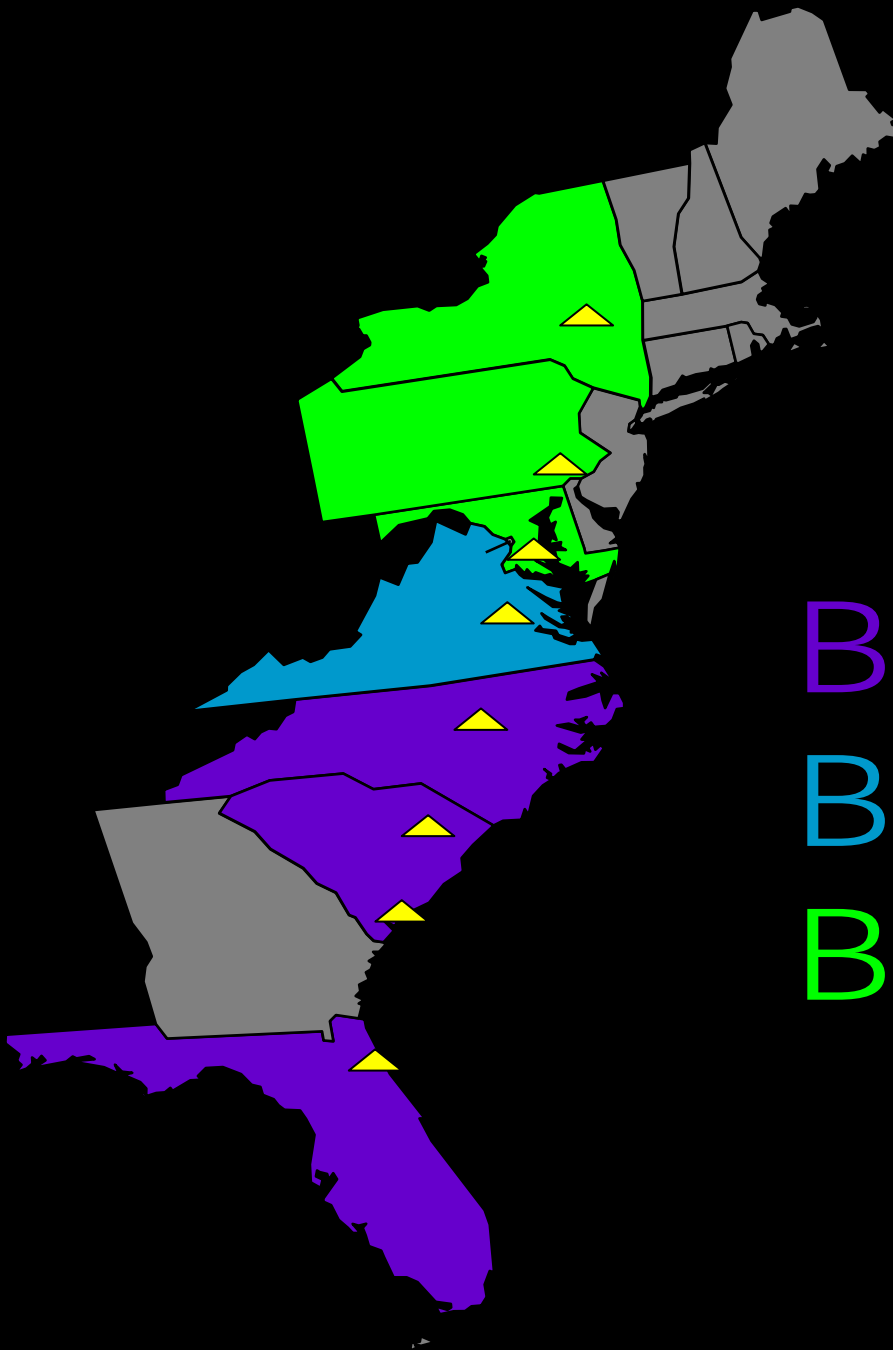
UPREGULATED GENES (OXIDASES, GSTs)

POINT MUTATIONS (KDR)

Culex spp. WGB19-95 est B Study

0.02 substitutions/site





B1, B2

B1, B2, B5

B1, B2, B5, BP

Analysis

Fit Points

Second Derivative Maximum

Baseline Adjustment

None

Arithmetic

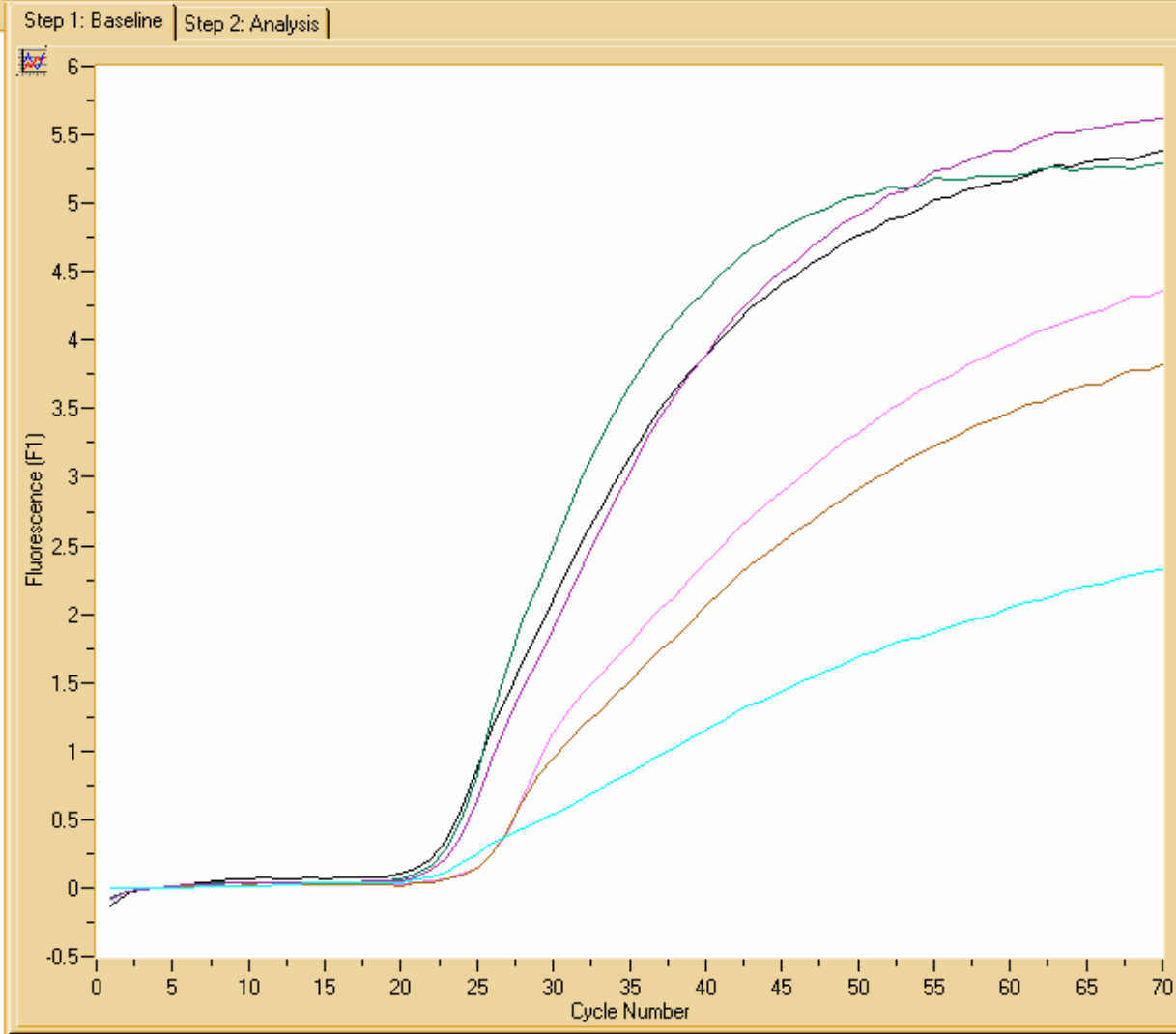
Proportional

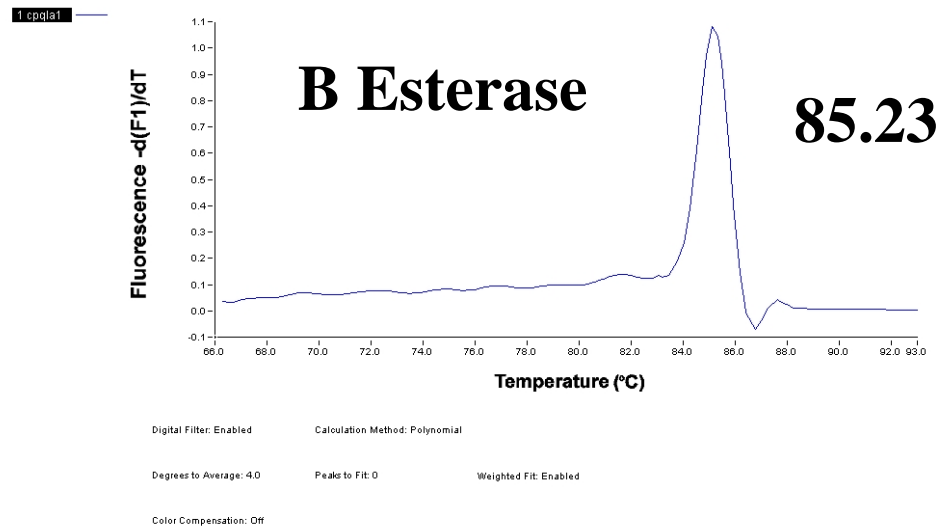
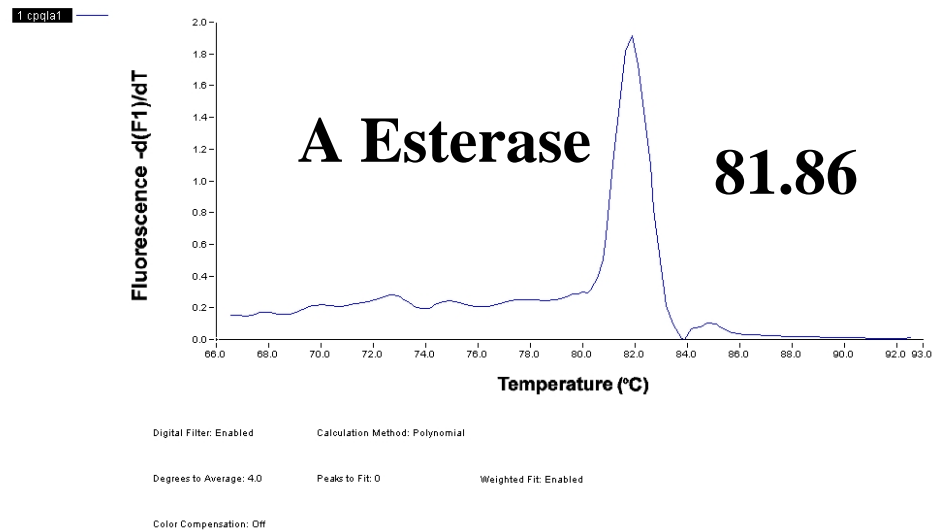
Normalized

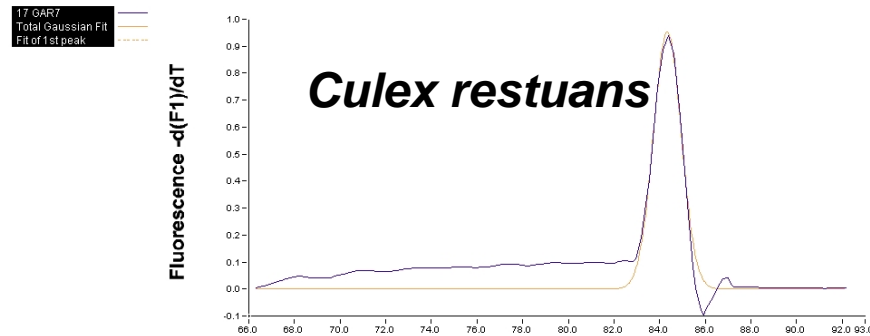
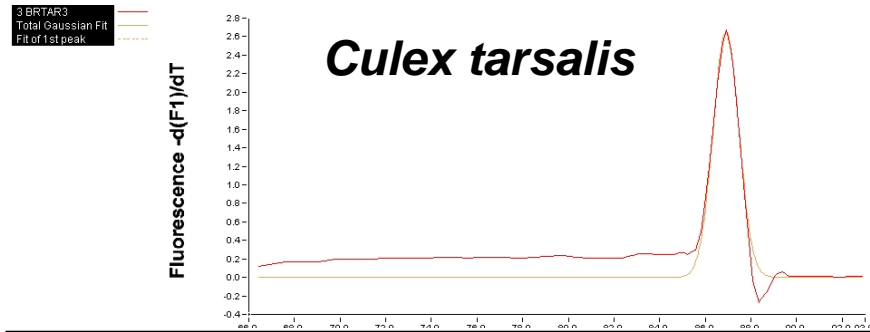
Analysis Notes

Empty text area for notes.

P...	Name	Calculat...	Cro...
1	BRTAR1	0.000E+00	22.76
2	BRTAR2	0.000E+00	23.20
3	BRTAR3	0.000E+00	23.06
4	BRTAR4	0.000E+00	22.13
5	BRTAR5	0.000E+00	23.49
6	BRTAR6	0.000E+00	22.83
7	BRTAR7	0.000E+00	23.14
8	BRTAR8	0.000E+00	23.49
9	BRTAR9	0.000E+00	22.87
10	BRTAR10	0.000E+00	22.79
11	CTL1	0.000E+00	22.94
12	CTL2	0.000E+00	22.49
13	CTL3	0.000E+00	22.89
14	CTL4	0.000E+00	23.28
15	CTL5	0.000E+00	25.24
16	CTL6	0.000E+00	24.10
17	CTL7		
18	CTL8	0.000E+00	21.29
19	CTL9	0.000E+00	23.17
20	CTL10	0.000E+00	24.71
21	NEG	0.000E+00	21.83







Temperature (°C)

Digital Filter: Enabled

Calculation Method: Polynomial

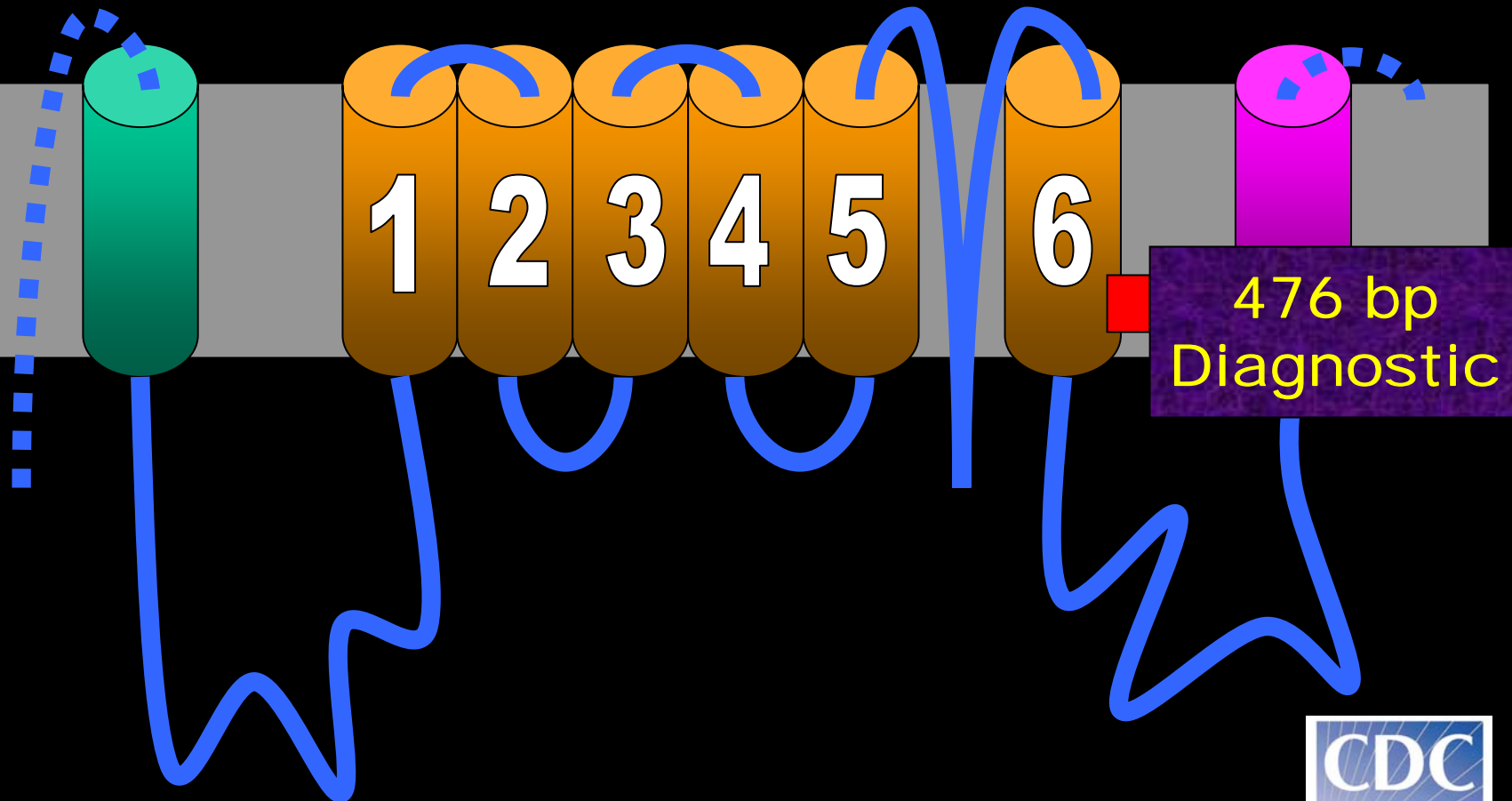
Degrees to Average: 40

Peak to Fit: 1

Color Compensation: Off

Esterase B

Diagnostic region for *Culex* point mutations in the sodium channel gene :

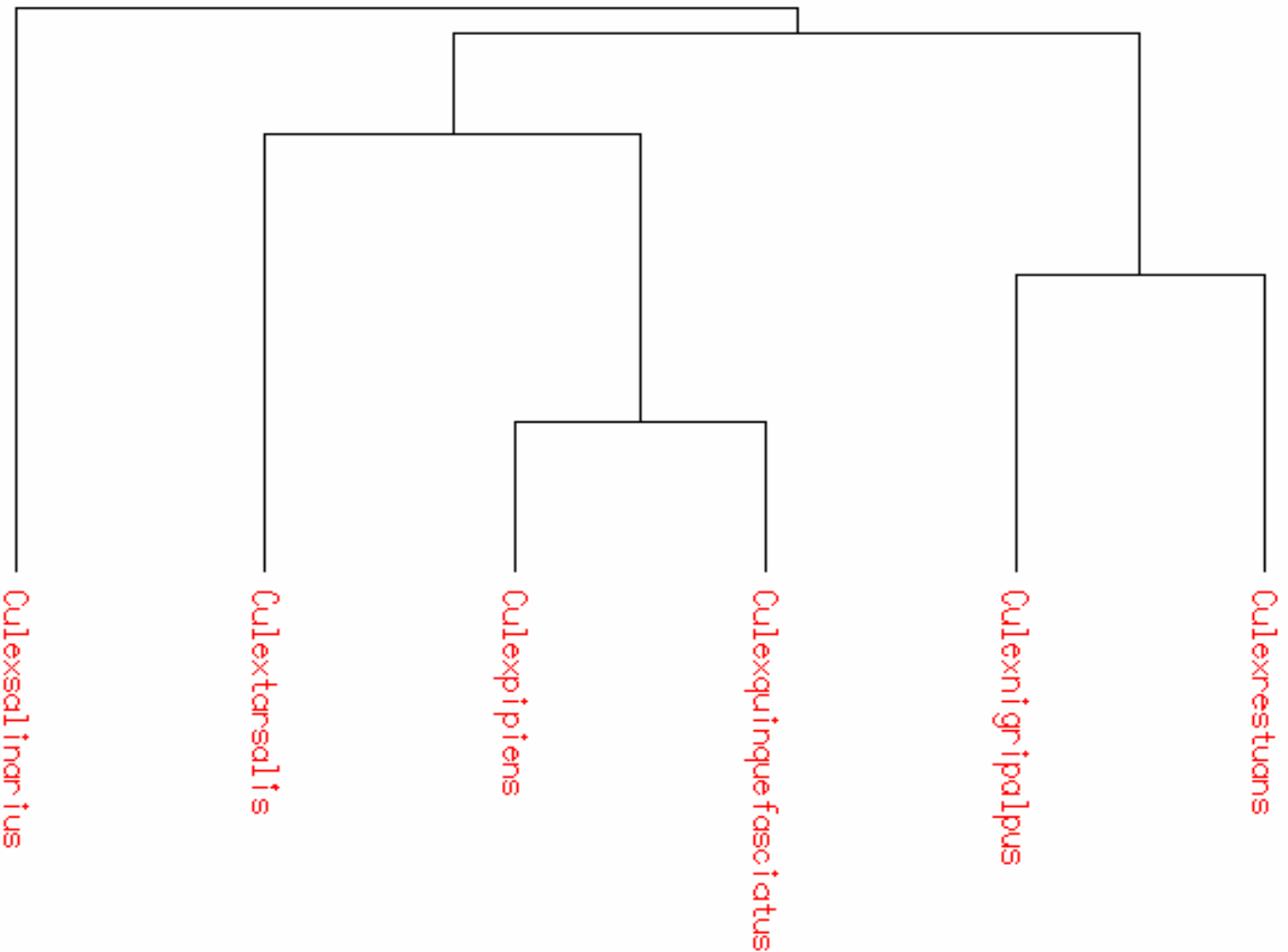


	1					50
res	TTAGTCGTGA	GTAT.....TCCAGCG..	TG
pip	TTAGTCGTGA	GTAT.....TCCAGTG..	TG
qnq	TTAGTCGTGA	GTAT.....TTCACC...	G
nig	TTAGTCGTAA	GTAA.....TCAAGC...	AG
sal	TTAGTCGTAA	GTATTGCTAG	TGGGGCCAAG	ATCCAACGGC	CCGGAAGAAG	

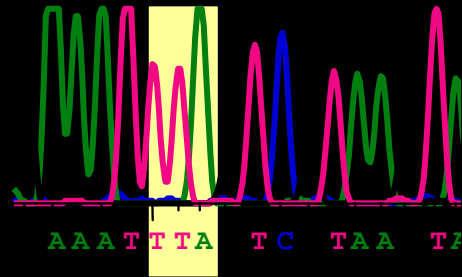
	51					100
CK30	AAGTCT....	.TCGTGATTG	A...TCTAGT	GTGCGCGCTA	GAGCTGTCAA	
CK02	AAATCT....	.GAGGGATTG	A...TCTAGT	GTGCACATTA	GAGCTGTCAA	
CK29	GAGTCT....	.TGGAAA...TGAACATAA	GAACTGTCAA	
CK13	GAATCT....	...GAGAT..	...TCAAGT	TTGTATAGTA	GAACTGTCAA	
CK09	GAGTCTGTAT	GGAGGGGGTG	AAATTCAAGT	GTGAATTGTT	GAACTGTCAA	

	101					150
CK30	AACATCGCCA	ACAGCATGCA	AGAAAAGGTG	GGAACGAAAA	ACTTTAAGGT	
CK02	AACATCGCCA	AGAGCATGCA	A.TAAAGGTA	AGCACGAAAA	ACTCTAAGGT	
CK29	AACATCGCCA	AGAGCATG.T	AGGAAAGGTG	GGAACGAAAA	ACTTTTAGGT	
CK13	AACAG.GTCA	A.AACTTCTG	AAATTCCGAA	ATTTCAAAAT	TCCAAAATTC	
CK09	AATAACGTCA	AGATTGTGTA	AG.AAAGGTG	AGC...AGAA	TCATGAAACT	

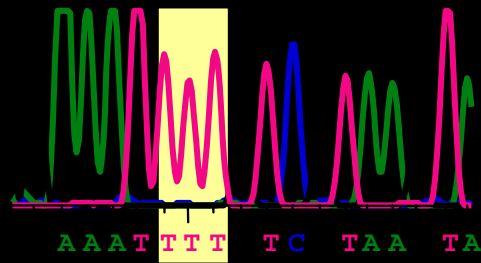
	151					193
CK30	CACATTTGTA	CCTTTGATGT	AAACAAACAG	TTC~~~~~	~~~	
CK02	AACATTTGCA	CCTTTGATGT	AAACAAACAG	TTC~~~~~	~~~	
CK29	AACATTTGAT	CTTTGGATGT	AAACAAACAG	TCA~~~~~	~~~	
CK13	CAAAATTCCA	AAATTCCAAA	ATTCCAAAAT	TCCAAAATTC	CAA	
CK09	CGCA~~~~~	~~~~~	~~~~~	~~~~~	~~~	



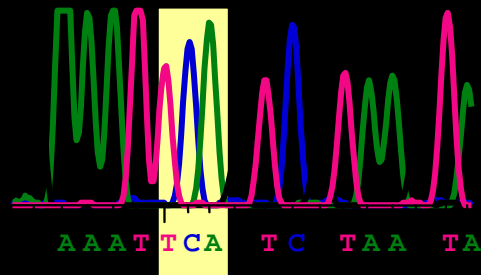
Leucine TTA - S

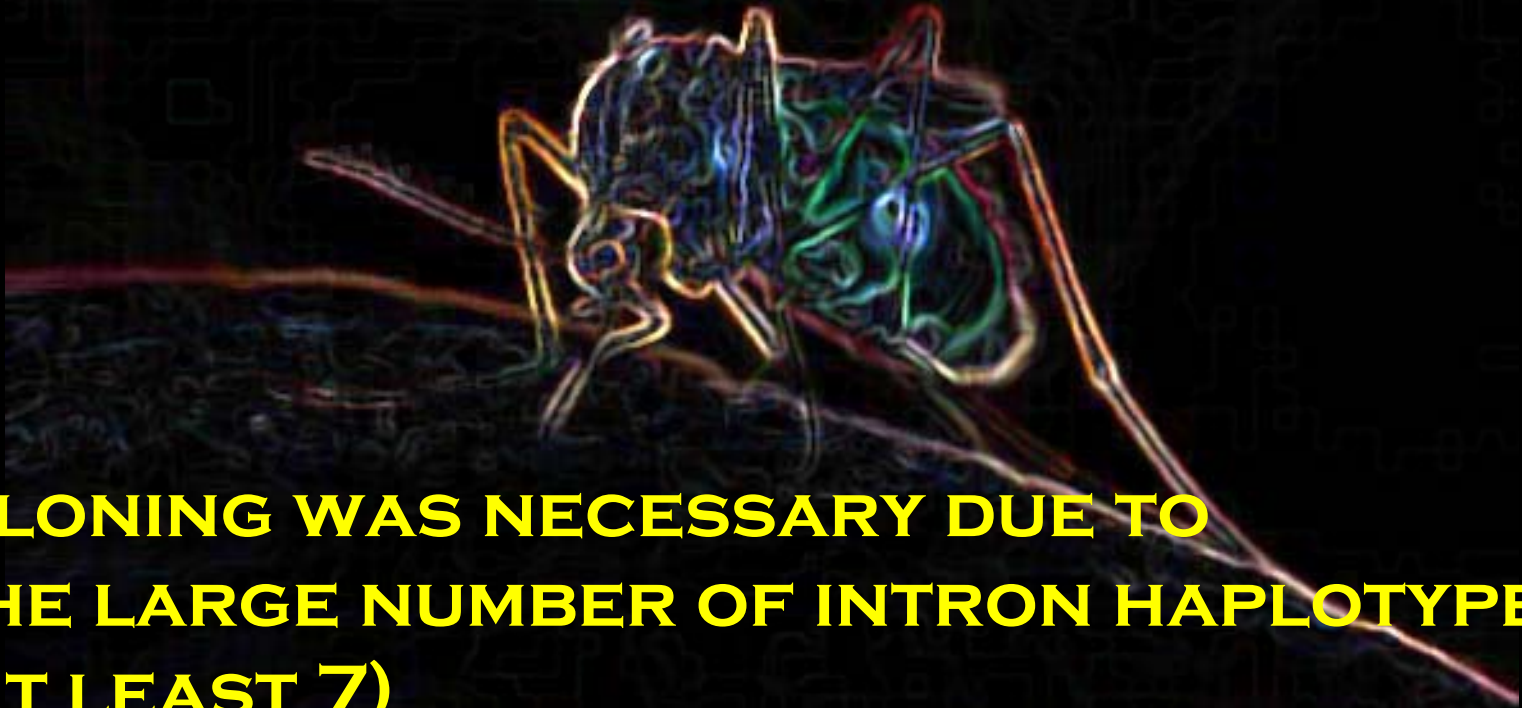


Phenylalanine TTT - R1



Serine TCA - R2





**CLONING WAS NECESSARY DUE TO
THE LARGE NUMBER OF INTRON HAPLOTYPES.
(AT LEAST 7)**

**THE KDR MUTATIONS IN DIFFERENT
POPULATIONS ARE ASSOCIATED WITH
DIFFERENT INTRON HAPLOTYPES.**

-TAGCGA-

-TCGTGA-

-TTTTAA-

-AAGAGA-

-TGGAAA-

-GAGGGA-

-TTGAGT-

-TAGCGA-

TTT

TCA

ALBANY

100%

100%

COLUMBIA

100%

→

JACKSONVILLE

20%

90%

-TCGTGA-

ALBANY

0%

0%

COLUMBIA

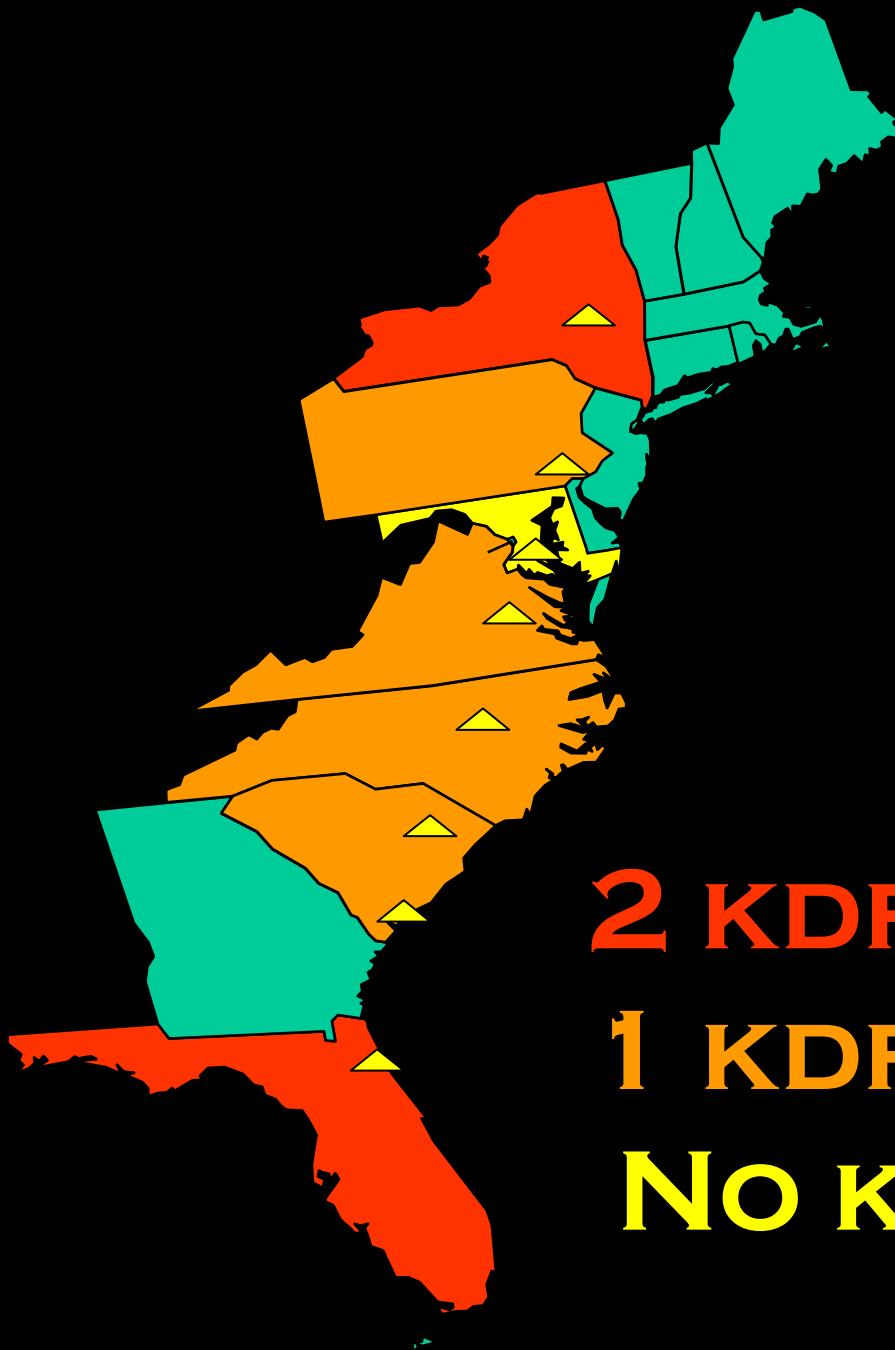
0%

→

JACKSONVILLE

80%

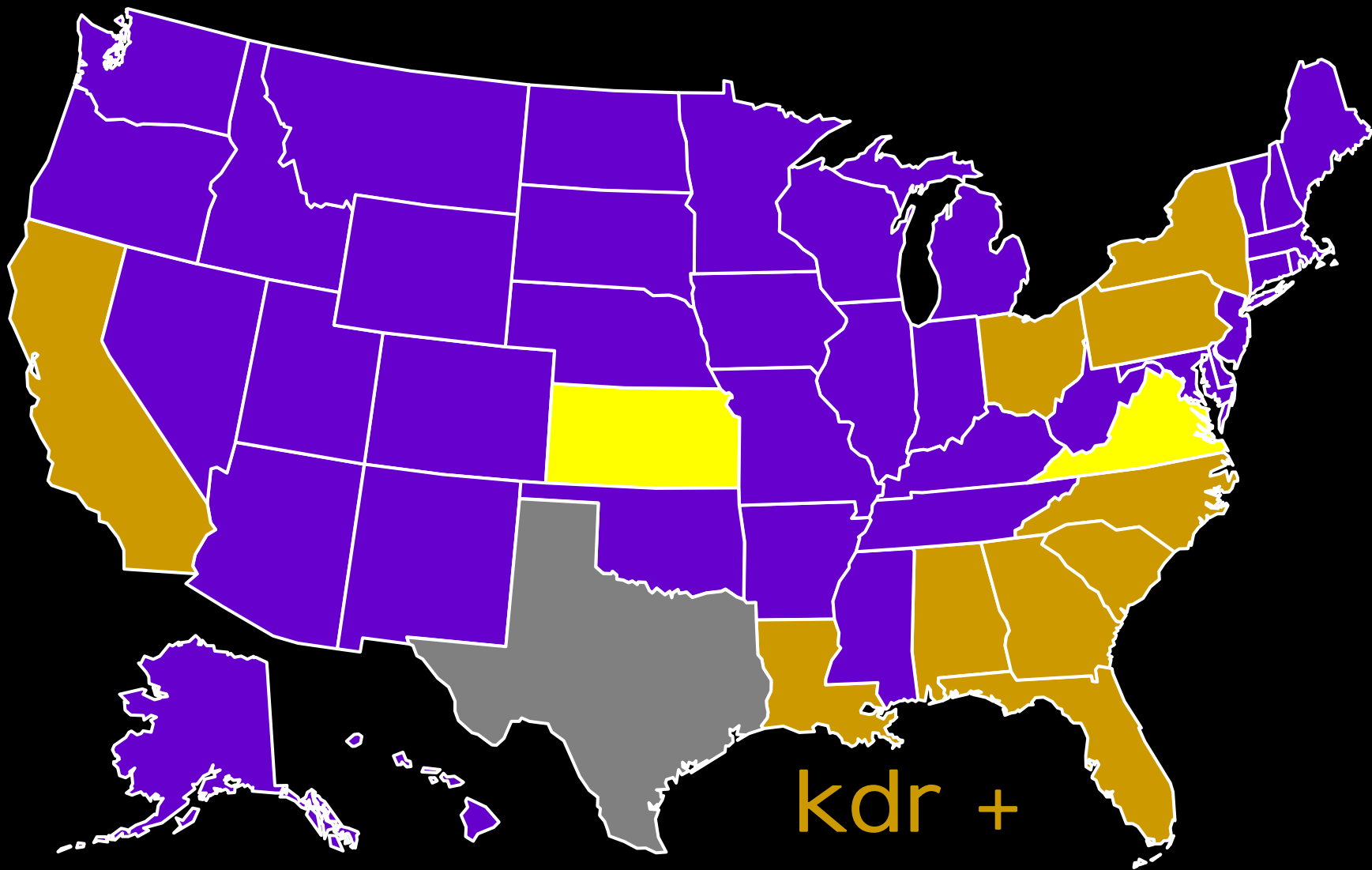
10%



2 KDR MECHANISMS

1 KDR MECHANISM

NO KDR



$kdr +$
 $kdr -$

UPREGULATED GENES

OXIDASES

GLUTATHIONE S-TRANSFERASES

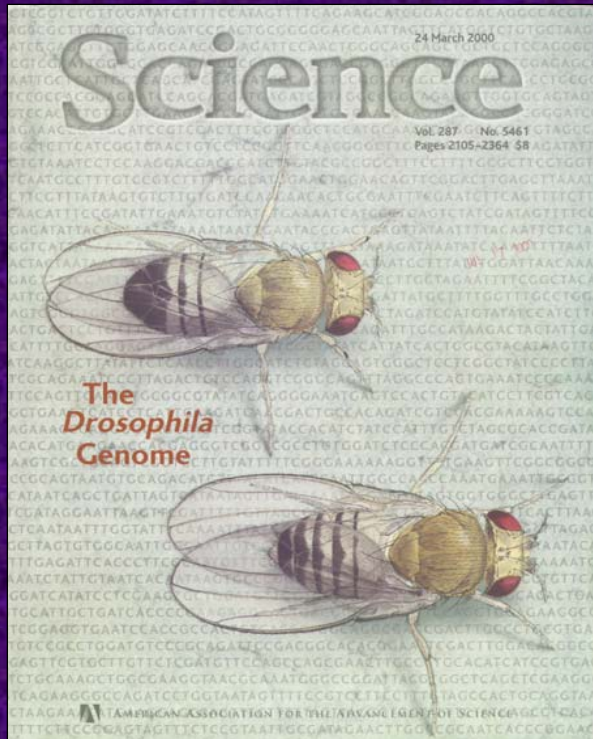


Enhanced oxidase Activity



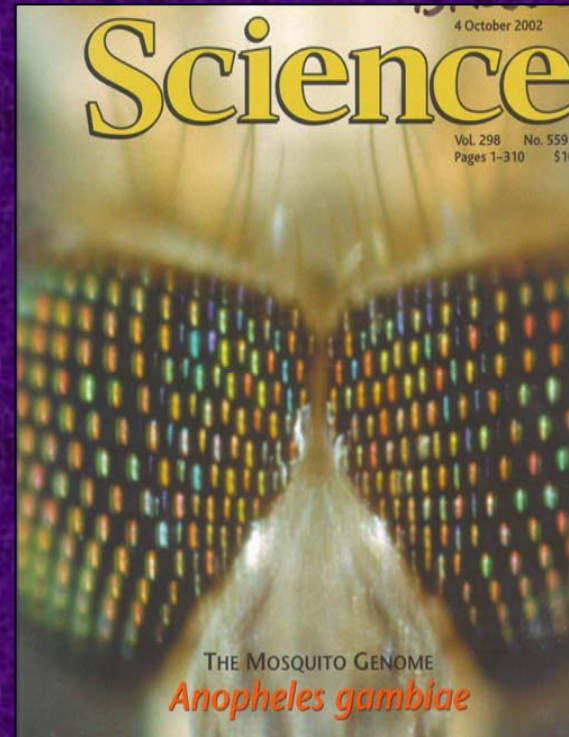
P450 Oxidase

Polymorphism



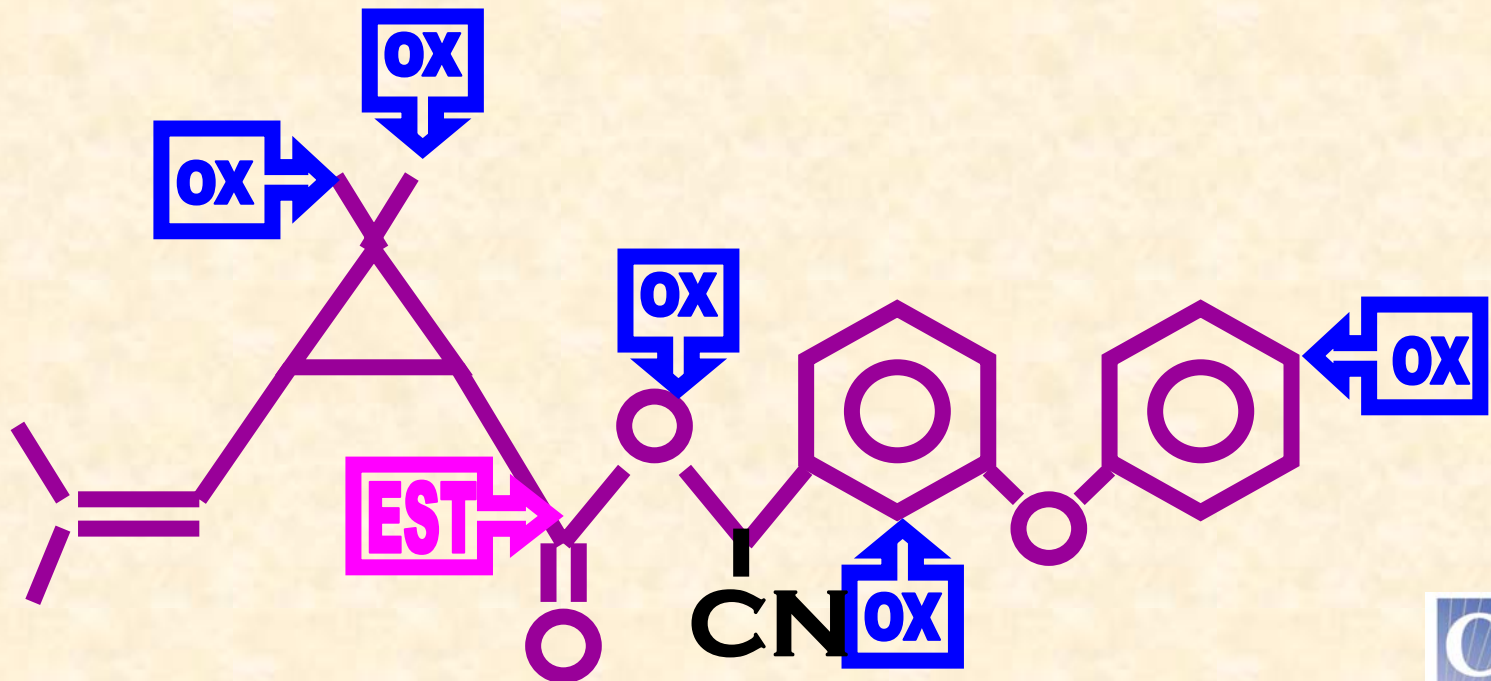
Drosophila
80 P450 genes

Anopheles gambiae
111 P450 genes



***Culex pipiens* – 52 P450 genes thus far.**

Detoxification enzyme attack on a synthetic pyrethroid:



1 MFAWIICAAA AVPLVYFLIV YQFSYWKRRG ITQLTPSFPP GDLGPFFRQR
51 SSLGVVYADV YRLCKRLPFV GIYLSLRPML VVNDPELIKN VLVRDFDHFH
101 DRGLYVNEEK DPLSGHLFAL GGEQWRHRS KLTPTFTSGR LKEMFTNLVQ
151 IGRVLQDHVA KRAGEDIEIR DVMARYTTDI IASVGFGIEN DSINEKGNIF
201 REMGTKVFSP DLKTILRLTS TFFTPKLNAL FGFKFIAQEI EDFIMNVVRE
251 TLEYRESNKV VRKDMMQLLM QLRNSGTVSI DDRWDIEVST NKKKLSLEQV
301 TAHAFVFFIA AYET SSTTIS FCLFELARNP EIQKKVQQEI DQVLASHNGE
351 ITYDNINEMK YLENCIDETL RKYPAVPFLN RECSKDYKIP GTDTTIEKGT
401 SLVIPVLGLH RDPDHYPEPD RFIPERFSNF EDISTKPYLP FGAGPRNCIG
451 LRLGKLQTKA GLVMMLSKFN VRLADETYAS KELALDARSV VLMPVGGIKV
501 SISERRAS*Y KLK*LVI*YL NITINDTKIN YL

Anopheles gambiae

D P E F Y P E P D Q F N P D R F M P E E
D P E Y Y P E P D Q F N P D R F L P E E
D P E F Y P E P D Q F D P D R F L P E E
D P E Y Y P E P E R F D P D R F L P E E
D P E F Y P E P D R F D P D R F L P E E
D P E Y Y P E P D R F D P D R F L P E E
D P E Y Y P E P D R F N P D R F L P E E
D P E Y Y P E P E R F D P E R F L P E E
D P E Y Y P E P D Q F D P D R F L P E E
D P D Y Y P E P D R F D P D R F V P E E
D Q E Y Y P E P D R F N P D R F L P E E
D Q E Y Y P D P E R F D P D R F L P E E
D P E Y Y P D P E R F D P D R F L P E E

Culex quinquefasciatus

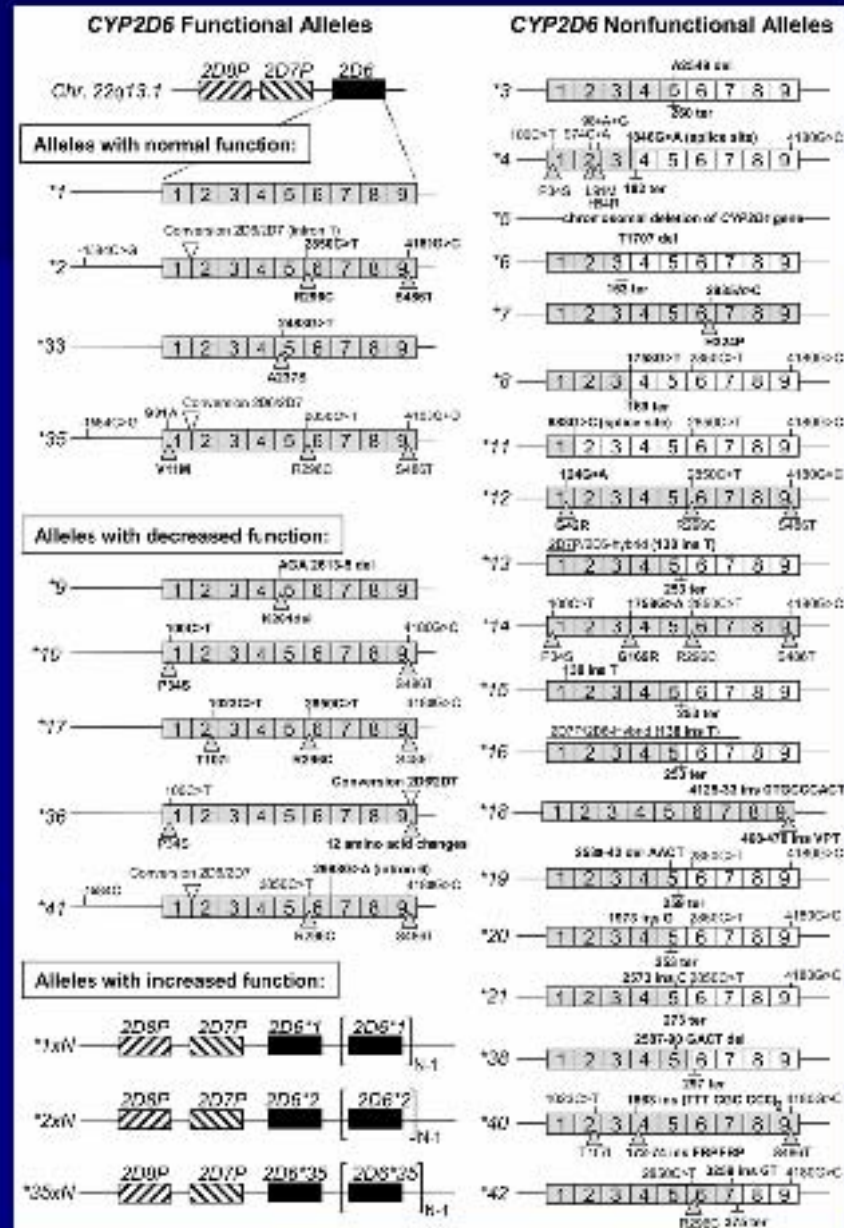
DPDHYPEPDRFIPERF SNFE
DPDIYPNPEVFNPERFIPEL

Culex pipiens

DPEQFPDPERFD PDRFLPEH
DPDQFPDPERFD PDRFLPE S

Alleles of *CYP2D6*

- More than 80 known
 - SNPs
 - INDELs
 - Complete gene deletion or duplications
- Clinical significance of most unclear



Cytochrome P450s (CYPs)

Genetic variants are associated with altered drug levels, but not with disease

- CYP2D6: estimated to metabolize 25% of all prescribed drugs
- CYP2C9: 5%
- CYP2C19: 15%
- CYP3A4/5: 50%



Examples of drugs metabolized by

CYP2D6

- Antiarrhythmics
- Antipsychotics
- Antidepressants
- Opiates
- Selective serotonin reuptake inhibitor (SSRI)

CYP2C9

- Anticonvulsants
- Antidepressants
- Hypoglycemics
- Anticoagulants
- Antibacterial
- Cancer Chemotherapy
- Nonsteroidal anti-inflammatory drugs

CYP2C19

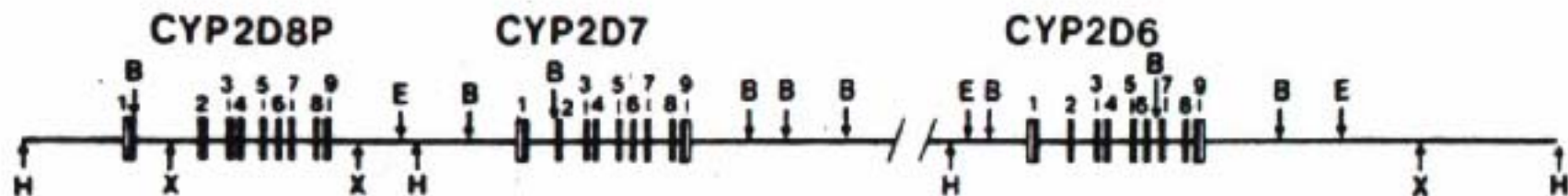
- Anticonvulsants
- Antidepressants
- Anti-ulcer
- Antimalaria
- Cancer Chemotherapy
- Proton pump inhibitors

Example:

Antidepressants and CYPs

Brand Name	Generic Name	Mechanism	CYP Metabolism
Effexor	Venlafaxine	<i>mixed</i>	CYP3A4, CYP2D6, CYP2C9, CYP2C19
Wellbutrin	Bupropion HCL	<i>mixed</i>	CYP2B6, CYP2D6, CYP1A2, CYP3A4 +
Desyrel	Trazodone	<i>mixed</i>	CYP3A4, CYP2D6
Zoloft	Sertraline	SSRI	CYP3A4, CYP2D6, CYP2C9, CYP2C19
Prozac	Fluoxetine	SSRI	CYP2B6, CYP2C9, CYP2C19, CYP3A4+
Celexa	Citalopram	SSRI	CYP3A4, CYP2C19, CYP2D6
Lexapro	Escitalopram	SSRI	CYP3A4, CYP2C19
Elavil	Amitriptyline	TCA	CYP3A4, CYP2D6, CYP2C19, CYP2B6+
Norpramin	Desipramine	TCA	CYP2D6, CYP1A2
Sinequan	Doxepine	TCA	CYP2D6, CYP1A2, CYP3A4
Pamelor	Nortriptyline	TCA	CYP1A2, CYP2D6, CYP2C19, CYP3A4
Anafranil	Clomipramine	TCA	CYP2D6, CYP1A2, CYP2C19, CYP3A4

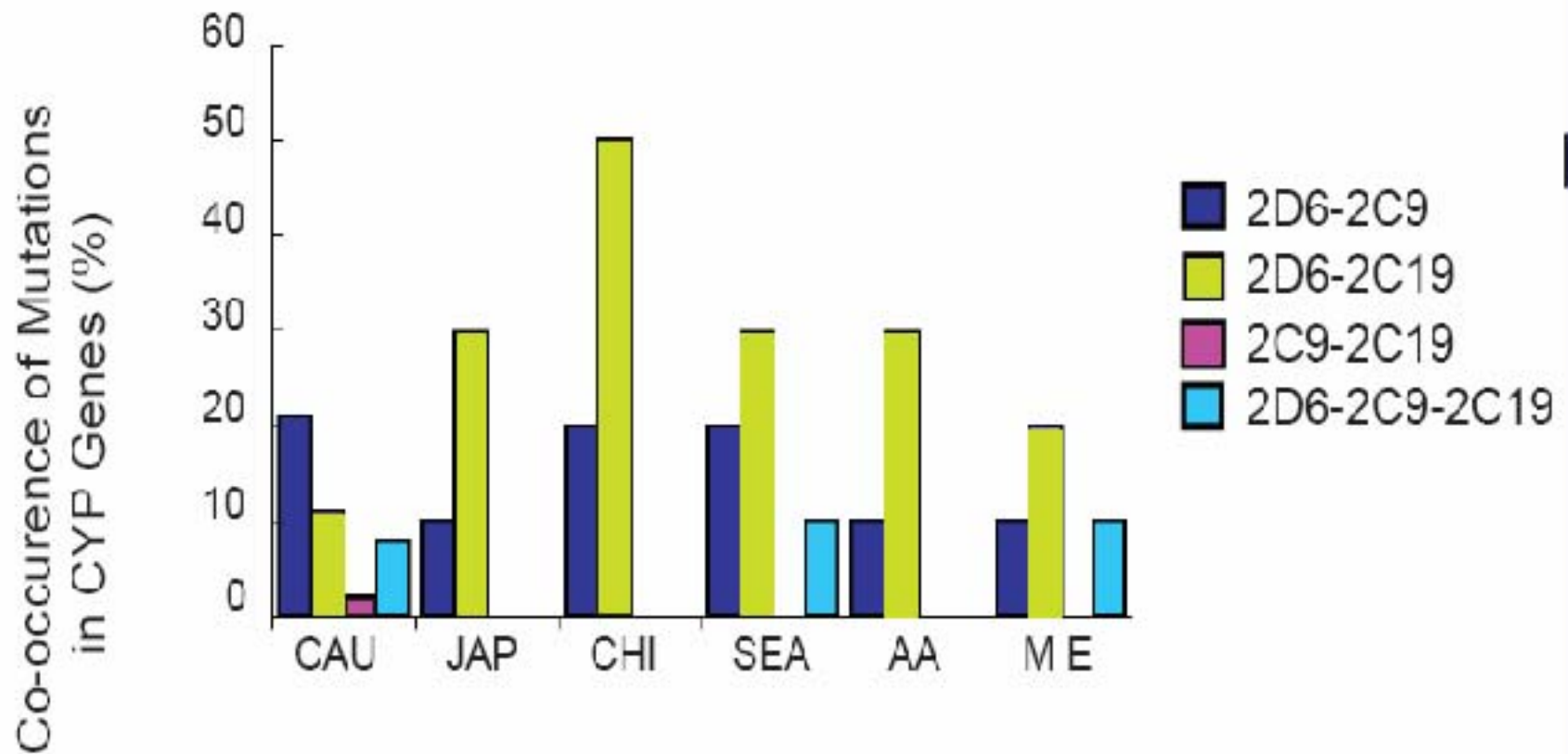
CYPs: Analytical challenges



Lancet 1990; 336: 529-32

- Superfamily/Pseudogenes:
 - 57 genes, 33 pseudogenes
- Gene dose important
- Many SNPs are common among alleles
- Many rare variants
- Importance of haplotypes not known

Co-occurrence of mutations in 2D6, 2C9, 2C19



GSTs

INSECT SPECIFIC

DELTA, EPSILON CLASSES

GSTE2 DDT, PERMETHRIN RESISTANCE IN *Aedes aegypti*

MORE WIDELY DISTRIBUTED

THETA, SIGMA CLASSES



QUESTIONS ?