The use of larvicides to control *Culex spp.* mosquitoes in catch basins

Why a presentation on larvicides?

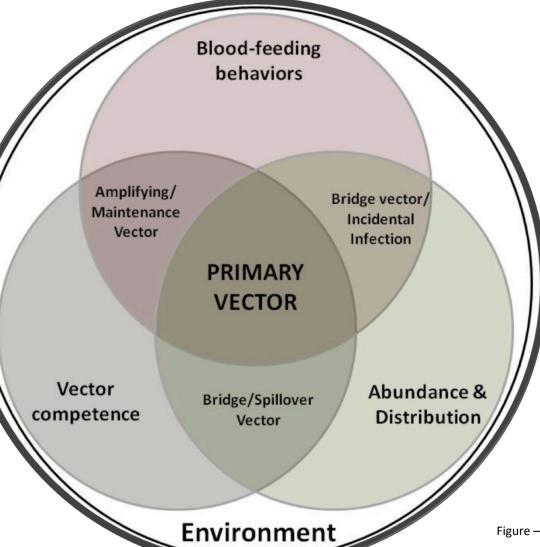
A pilot study related to my dissertation:

The amplification and transmission of West Nile virus in a two vector system: examining the role of secondary vectors

Chapter: Experimental perturbations of *Culex spp.* populations and the effect on WNV transmission

Qualification Exam 4/9/15

Defining primary and secondary vectors



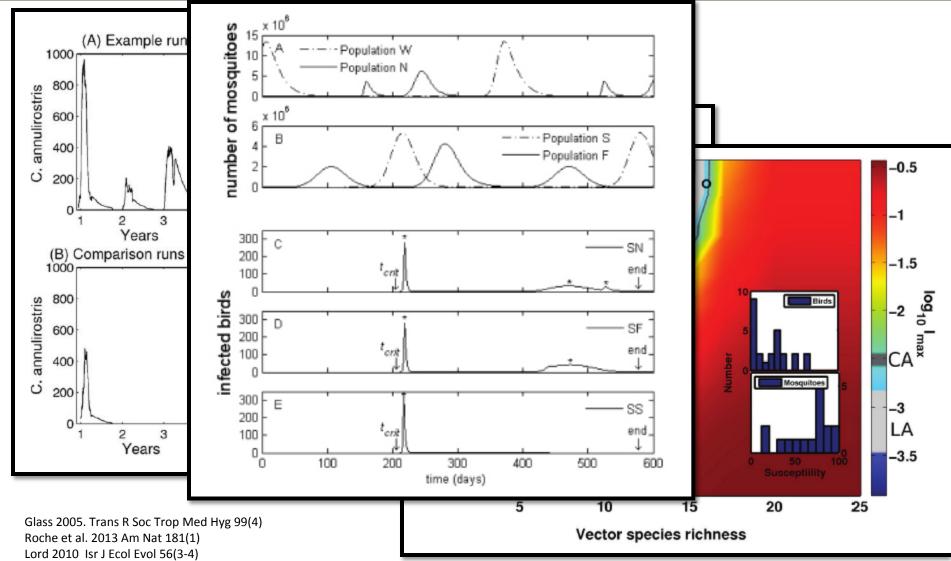
The distinction of a primary vector is often based on epidemiological rather than on ecological characteristics

<u>Public health perspective</u>: Primary vectors are those that cause the most human morbidity and mortality

Ecological perspective: Primary vectors are those most responsible for periods of peak transmission

Figure – McMillan PBEE Qualifying Exam Literature Review

Secondary vectors and vector-borne disease transmission



The primary and secondary vectors of West Nile Virus

The Big Dawgs

Members of the *Culex pipiens* species complex:

- Culex pipiens pipiens
- Culex quinquefasciatus
- Hybrids of the two subspecies and potentially the form *molestus*

The Other Guys

Native *Culex* species and perhaps a few non-native *Aedes*:

- Culex tarsalis
- Culex restuans
- Culex nigripalpus
- Aedes albopictus
- Ochlerotatus japonicus

Moving beyond models: experiments

We know...

- *Culex spp.* commonly breed in road-side catch basins
- Cx. restuans is active in spring/early summer
- *Cx. quinquefasciatus* dominates field collections during summer months
- Difficult to identify *Cx. restuans* and *Cx. quinquefasciatus* from a gravid trap

Thinking experimentally

Use catch basins as a unit of intervention

Apply larvicides during time periods that coincide with each vector species period of greatest activity

Monitor the effect of the insecticide application at the catch basin level

Monitor WNV infection in mosquitoes using gravid traps Individually vial any *Culex* spp. we cannot identify Monitor WNV infection in wild birds

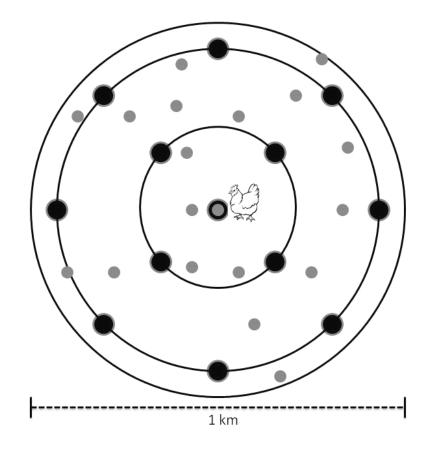
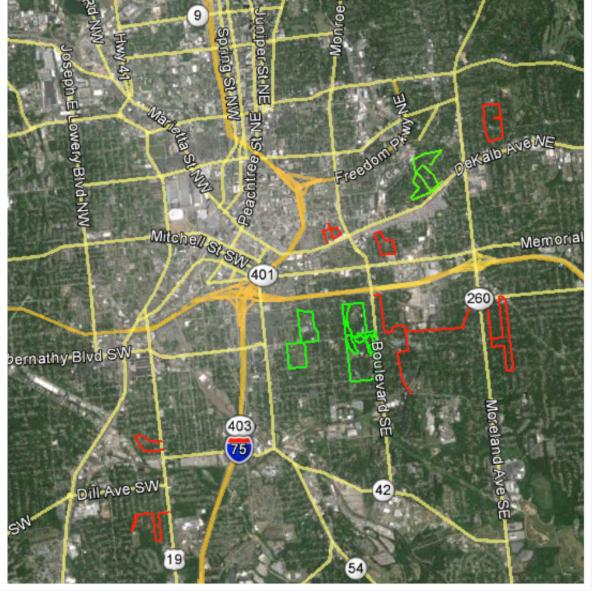


Figure – McMillan PBEE Qualifying Exam Proposal

SUMMER 2015 PILOT STUDY

Catch hasin explorations



ita, where are Culex breeding in catch **s**? red public parks and borhoods unding parks ked basins for: hter y presence of *Culex* squitoes

Paired Experimental Design

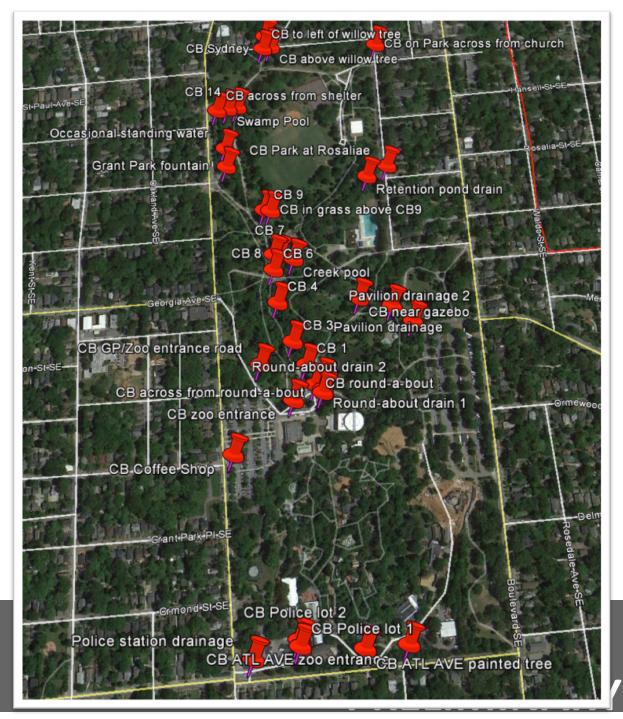
- Sampled two parks (w/ 1 mile of each other)
 - Grant Park (treatment park) 7 catch basins
 - Phoenix Park (control park) 10 catch basins
- Catch basin surveillance:
 - 3 dips for egg, larvae, and pupal collections
 - 5 minutes aspiration for adult mosquito collections
 - Measurement of basin water depth
 - Water quality sample for ammonia, nitrate, and phosphate measurements
 - All IV larvae, pupae, and adult identified to species

Larvicide applications

- 36 permanent and semi-permanent water holding containers in Grant Park
- Treat weekly with Summit Chemical's Mosquito Bits and Mosquito Dunks
 - Bascillus thurenginsis larvicide
 - Bits (10.96% Bti) fast acting
 - Dunks (2.96% Bti) slow release
 - Approx. 3 tablespoons of bits and up to 2 dunks applied per week
- Application period: July 16th September 9th 2015

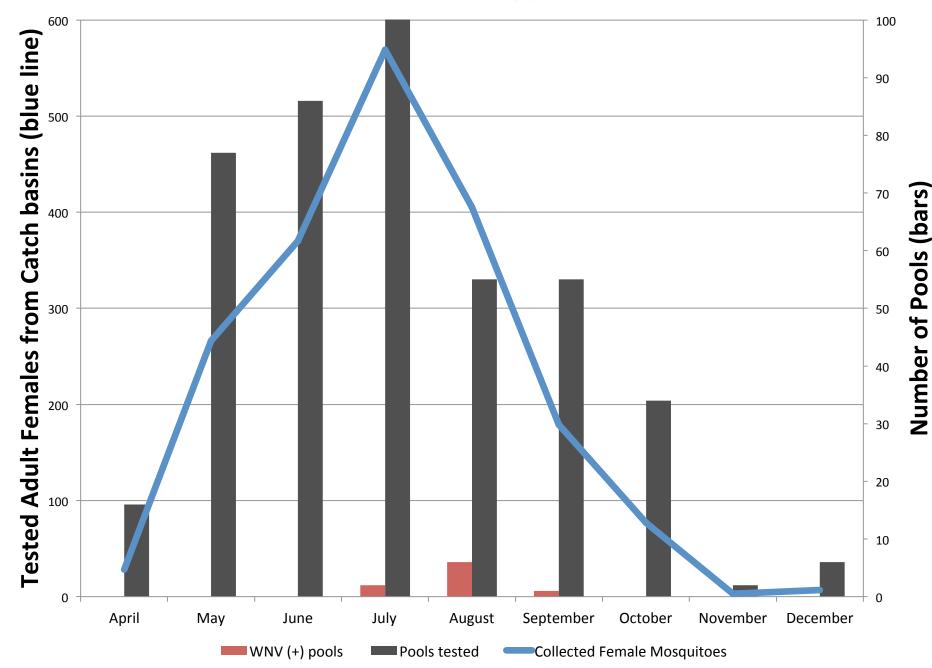
WNV Surveillance

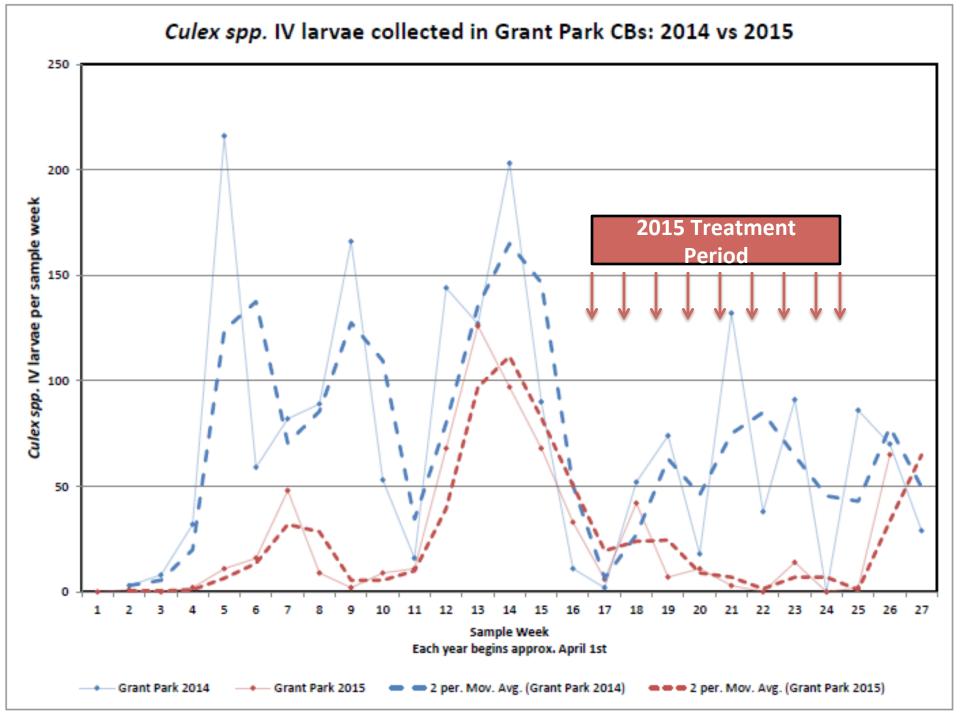
- Gravid trap surveillance
 - Weekly in both parks
- Light trap surveillance in sewers (Grant Park only)
 - Weather permitting
- All adult female mosquitoes pooled by site, collection method, date, and species
 - Unidentified *Culex spp.* pooled individually
 - Bloodfed mosquitoes pooled individually
- Mist netting for wild birds (Grant Park only)
 - Individuals age, sexed, banded
 - Blood sample taken from jugular vein

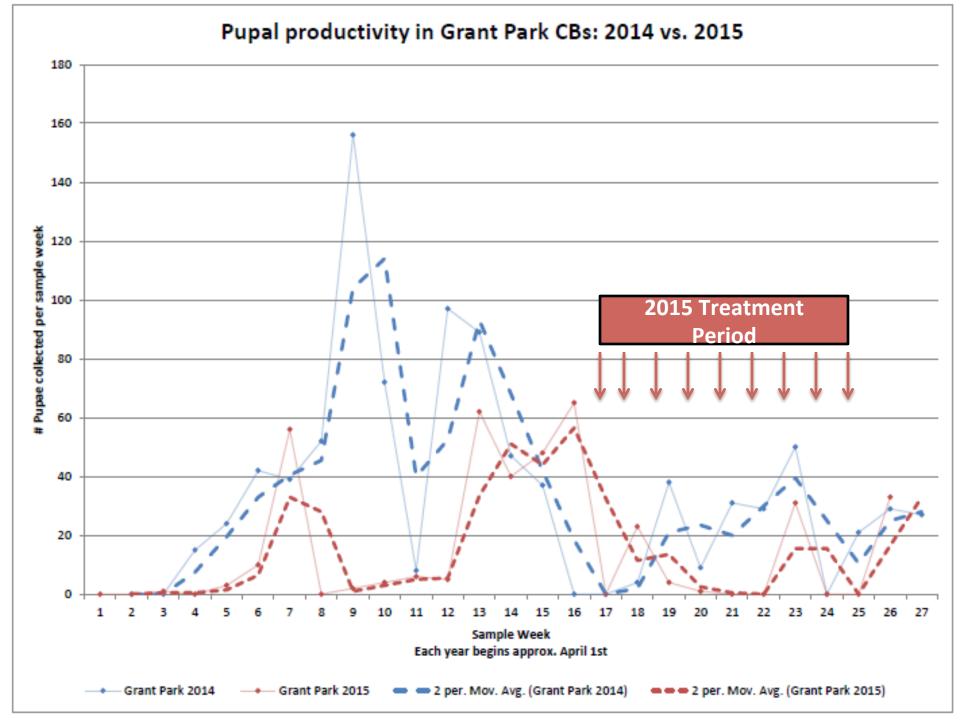


RESULTS

2014 Grant Park Adult Female Culex spp. collections in catch basins

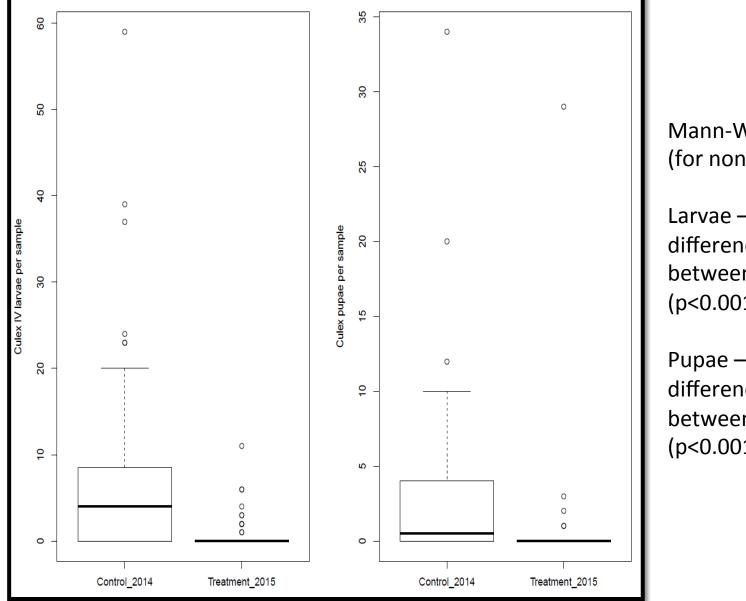






Culex spp. larvae

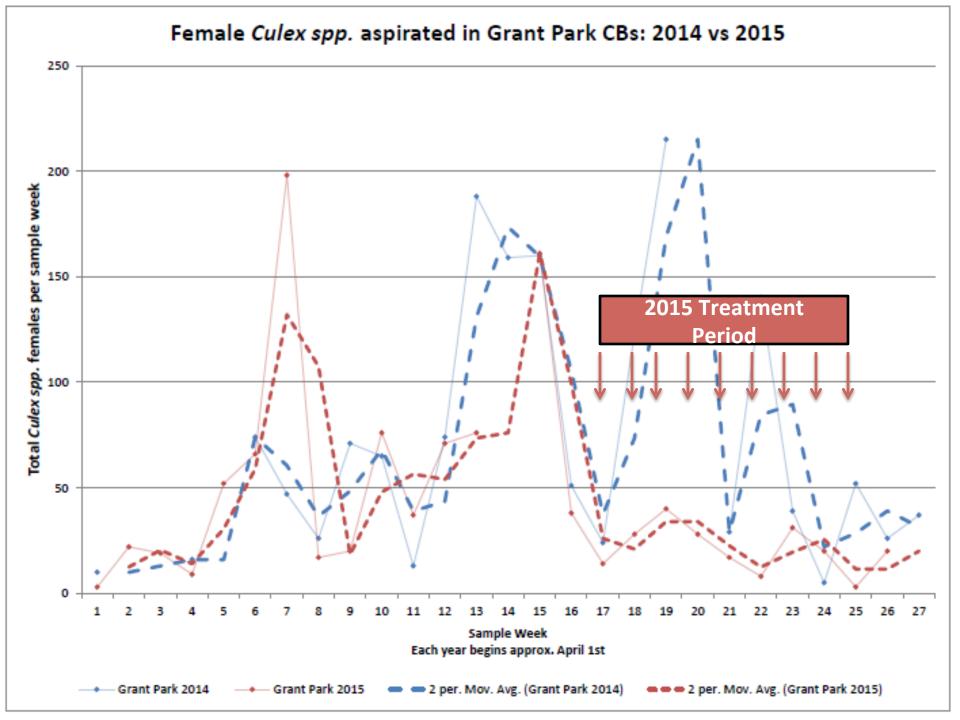
Culex spp. pupae

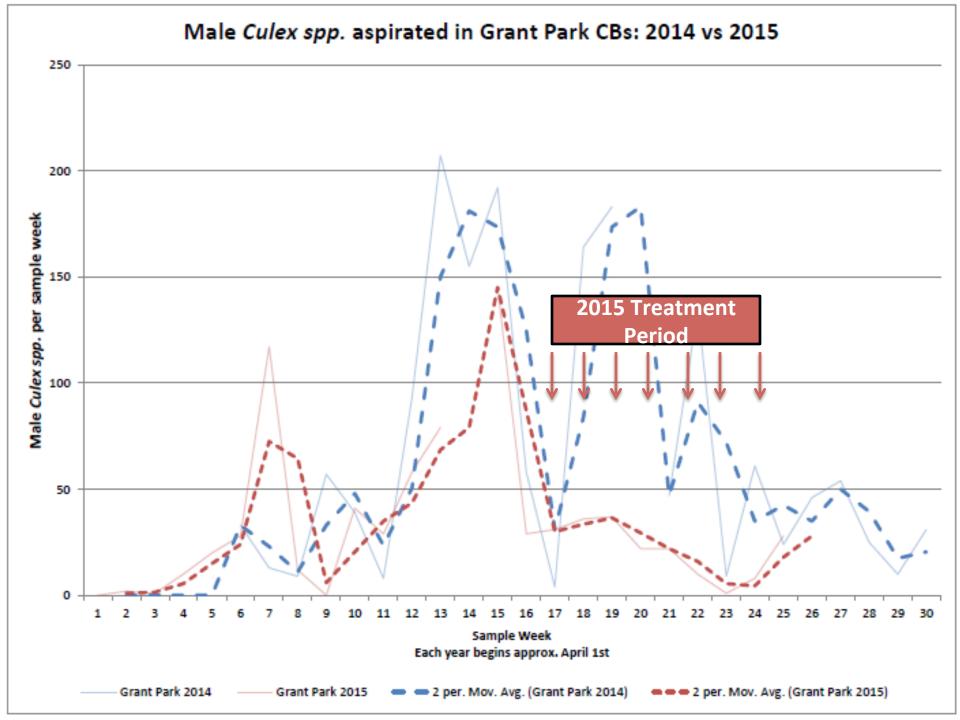


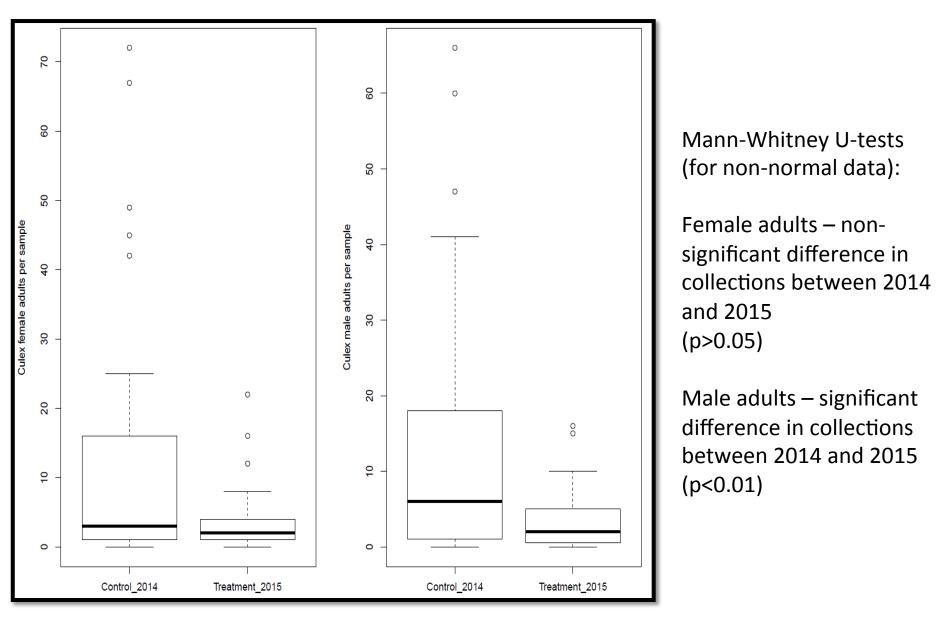
Mann-Whitney U-tests (for non-normal data):

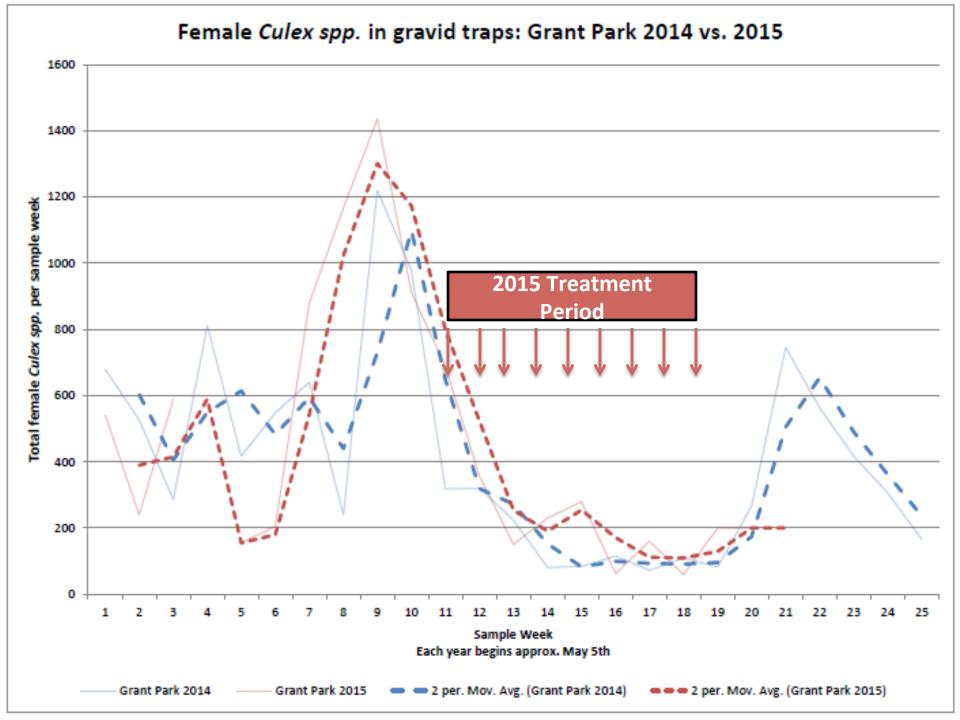
Larvae – significant difference in collections between 2014 and 2015 (p<0.001)

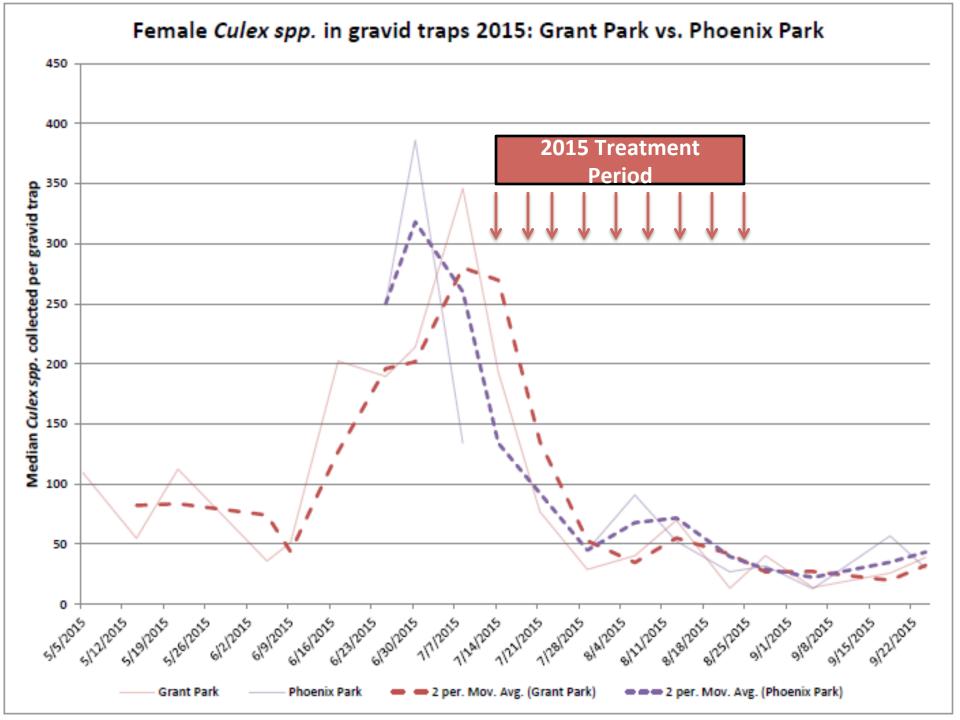
Pupae – significant difference in collections between 2014 and 2015 (p<0.001)

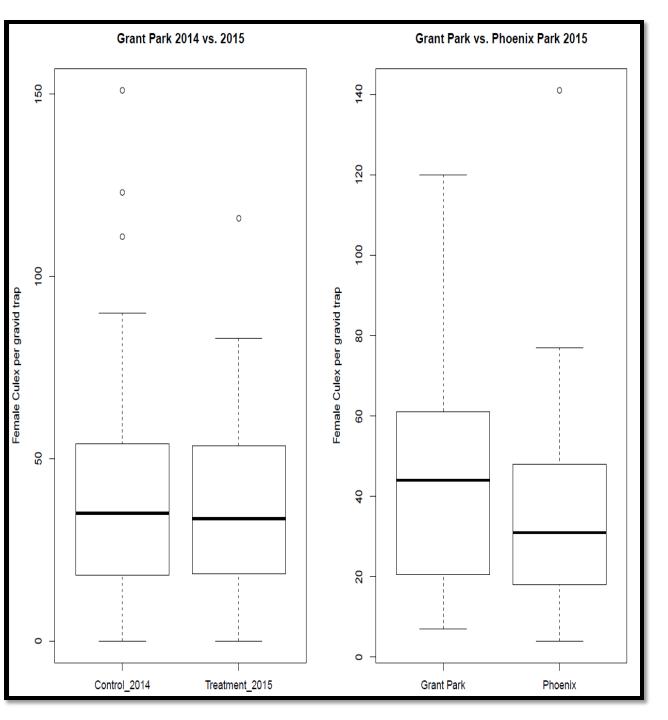












Mann-Whitney U-tests (for non-normal data):

Grant Park: non-significant difference in collections between 2014 and 2015 (p>0.8)

Summer 2015 paired EXP: non-significant difference in collections between Grant Park and Phoenix (p>0.25)

Bird data Summer 2015

| Species | # Blood Samples |
|----------------------|-----------------|
| American Robin | 18 |
| Brown Thrasher | 6 |
| Brown-headed Cowbird | 1 |
| Carolina Wren | 6 |
| Common Grackle | 2 |
| Downy Woodpecker | 1 |
| Eastern Bluebird | 5 |
| Eastern Phoebe | 1 |
| Eastern Towhee | 3 |
| Gray Catbird | 10 |
| Gray-cheeked Thrush | 1 |
| Northern Cardinal | 6 |
| Northern Mockingbird | 5 |
| Red-eyed Vireo | 1 |
| Song Sparrow | 2 |
| Swainson's Thrush | 5 |
| Yellow-breasted Chat | 1 |

- Blood samples from Grant Park ONLY
- 17 species sampled
- 76 total samples (as of 10/8/15)
- Success Rate: 63.7%
- 25 individuals bleeding not attempted

The data to come...

•WNV infection data for: •All mosquito pools •All avian samples Mosquito species identification for: •Sub-sample of individual Culex spp. pools Blood source identification for blood feds •Species identification for *Culex spp.* blood feds •Models that control for: •Weather (temperature/ precipitation) •Time-lags Insecticide application •Extreme weather events

Future Directions and Questions

Given the research question, are larvicides the most appropriate control tool?

At the catch basins level: residual spraying vs. larvicide vs. BOTH

How large is the above ground *Culex spp.* populations? Why are 2014 and 2015 gravid trap collections almost identical?

How large is the underground *Culex spp.* population? Are above ground and catch basin interventions impacting below ground populations?

Acknowledgments

- Advisor and mentor: Gonzalo Vazquez Prokopec
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THANK YOU & QUESTIONS?